MANIPAL ACADEMY OF HIGHER EDUCATION

MSc Clinical Virology Curriculum Manipal Institute of Virology, MAHE, Manipal

Dr. Chiranjay Mukhopadhyay, Director, MIV Manipal

2022

Outcomes Based Education (OBE) Framework

Two Year Full Time Post Graduate Program

MANIPAL INSTITUTE OF VIROLOGY, MAHE, MANIPAL-576104



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1. NATURE AND EXTENT OF THE PROGRAM

M.Sc. (Clinical Virology) Degree Programme

The two-year MSc Clinical Virology program is structured in four semesters for effective theoretical and practical learning. The entire program is of 80 credits. The program awards a degree with an international acclaim and world-wide recognition.

Duration of the Programme

The programme is of four semesters. Duration of each semester is six months. Each semester is composed of a set of courses and each course depending on the nature and scope of the subject consists of Lectures/Tutorial/Practicals. The student has to carry out a project work in the fourth and final semester of the programme (6 months).

Medium

The medium of instruction and examination is English.

Eligibility

Bachelor's degree in Life Sciences (Microbiology / Biotechnology / Biochemistry / Botany / Zoology) or MBBS / BVSc / BSc-MLT or any other related subjects from a recognized University, with minimum 60% aggregate marks or an equivalent CGPA.

MSc Clinical Virology programme aims at training students in the state of the art virological techniques useful in health, diagnostics, industrial, and academic sectors. Concepts of biosafety practices, outbreak investigations, clinical virology, etc. are also introduced, updated and strengthened through this programme. Students are practically trained to operate high end laboratory equipment for diagnostic and research work. Observation, communication, analysing information, problem-solving, critical thinking, logical reasoning, and perseverance are a few soft skills inculcated in students during the programme. The curriculum content involves extensive clinical, diagnostic, and laboratory practices in infectious diseases with virological aetiology. Special emphasis is laid on identifying and predicting future technological developments, changes in diagnostic service delivery and future patient/clinician/hospital requirements, to ensure that the curriculum is as robust and sustainable as possible. This is in relation to both scientific content and anticipated future developments and is reflective of the requirements of a specialized postgraduate program.



2. PROGRAM EDUCATION OBJECTIVE (PEO)

The overall objectives for M.Sc. Clinical Virology program are as follows.

PEO No.	Education Objectives
PEO 1	Students will be trained in syndromic approach of viral disease diagnosis with special emphasis on molecular and serological techniques.
PEO 2	Students will acquire fundamental and practical knowledge in subjects such as cell biology, molecular virology, virological techniques, biosafety and biosecurity, immunology, data analysis, disaster management, epidemiology and public health.
PEO 3	Students will learn and reflect ethical attitude, strong communication, and effective interpersonal skills in their professional practices and would be able to work cohesively in a team with multidisciplinary backgrounds.
PEO 4	Students will learn the importance of bioethics, research, innovation, and intellectual property rights in research.
PEO 5	Students will be able to achieve professional excellence by using their theoretical and technical competence in virology.
PEO 6	Students will be able to participate in lifelong learning process for a highly productive career and will be able to relate the concepts of virology in disease diagnosis and designing therapeutic strategies.



3. GRADUATE ATTRIBUTES

S. No.	Attribute	Description
1	Disciplinary Knowledge	Knowledge of all aspects of virology involving theoretical and practical techniques and other related areas of studies.
2	Understanding different subsets of Virology	Different areas of virology including, molecular virology, cell biology, bioinformatic, epidemiology, biosafety and biosecurity, biostatistics, and bioethics.
3	Measurable Skills and Industry-ready Professionals	Strengthening skills and knowledge regarding current updates in virological research and development in industries and research organizations.
4	Effective and Influencing communication	Efficient in sharing thoughts, ideas and applied skills of communication in various forms such as written and verbal communication.
5	Cooperation/Teamwork	Ability to work in teams as well as independently.
6	Critical/ Reflective thinking & language efficiency	Ability to employ critical and reflective thinking in diagnosing viral infections.
7	Technologically Efficient Professional	Capability to work with advanced techniques and high-end instruments used in diagnosis and research.
8	Research-related Skills	Trained to address research questions through short-term scientific projects.



4. QUALIFICATION DESCRIPTORS

1. Demonstrate

- (i) a systematic, extensive and coherent knowledge and understanding of virology, related disciplinary areas/subjects, and applications; including a critical understanding of the established theories, principles and concepts, and number of advanced and emerging issues in the field;
- (ii) procedural knowledge that creates different types of professionals related to virology, including research and development, teaching, government and public service;
- (iii) professional and communication skills.
- 2. Demonstrate comprehensive knowledge about current research, scholarly, and/or professional literature, relating to essential and advanced learning areas pertaining to virological techniques and skills required for identifying problems and related issues.
- 3. Demonstrate skills in identifying information needs, collection of relevant quantitative and/or qualitative data drawing on a wide range of sources, analysis and interpretation of data using methodologies as appropriate to the course(s) for formulating evidence-based solutions and arguments.
- 4. Use knowledge, understanding and skills for critical assessment of a wide range of ideas and complex problems and issues relating to virology.
- 5. Communicate the results of studies undertaken in an academic field accurately in a range of different contexts using the main concepts and techniques of virological studies.
- 6. Address one's own learning needs relating to current and emerging areas of study, making use of research, development and professional materials as appropriate, including those related to new frontiers of knowledge.
- 7. Develop the clinical, scientific, technical, management, communication and leadership skills required to run a diagnostic/research laboratory and deliver a high-quality clinical service.
- 8. Apply one's disciplinary knowledge and transferable skills to new/unfamiliar contexts and to identify and analyze problems and issues and seek solutions to real-life problems.



5. PROGRAM OUTCOMES

After successful completion of M.Sc. Clinical Virology program, students will be able to,

РО	Attribute	Competency
PO1	Disciplinary knowledge	Demonstrate comprehensive knowledge and understanding of courses that form a part of the postgraduate programme.
PO2	Communication Skills	Express thoughts and ideas effectively through written and verbal communication; establish communication link with others using appropriate media; share and express personal views confidently; reflect a good listener's trait; read and write analytically; process complex information and present it in a clear and concise manner.
PO3	Critical thinking	Apply a critical thinking process of identifying, analysing and reviewing clinical cases and demonstrate skills in mapping disease diagnosis algorithms.
PO4	Problem solving	Extrapolate from practical trouble-shooting experiences and apply the knowledge in solving various non-familiar problems.
PO5	Analytical reasoning	Evaluate the reliability and relevance of evidence; identify logical flaws and gaps in arguments; analyse and synthesise data from a variety of sources; draw valid conclusions and support them with evidence and examples, and address opposing viewpoints.
PO6	Research-related skills	Inquisitiveness to ask appropriate/relevant questions; ability to recognise and predict cause-and-effect relationships, define problems, formulate hypotheses, test hypotheses, analyse, interpret and draw conclusions from data; ability to plan, execute and report the results of an experiment or investigation. Ability to work in laboratory culture, learn to work independently and get exposure to scientific writing and publication through six-month dedicated research projects.
PO7	Cooperation/ Teamwork	Demonstrate leadership and teamwork with a positive attitude to effectively manage human resources.
PO8	Scientific reasoning	Ability to analyse, interpret and draw conclusions from quantitative/qualitative data; critically evaluate ideas, evidence and experiences through an open-minded and reasoned perspective.
PO9	Reflective thinking	Integrate theory and practice to develop work habits and attitude necessary for job success through practice school and professional events.



Moral and ethical awareness/reasoning Lifelong learning	necessary for continuous learning; use fundamental
awareness/reasoning	professional situations. Investigate and provide independent learning skills
Lifelong learning	necessary for continuous learning; use fundamental
	achieve professional excellence.
Multicultural competence	Sensitively react towards values and beliefs of different cultures, effectively engage in a multicultural society and interact respectfully with diverse groups across the globe.



6. COURSE CURRICULUM AND STRUCTURE

Course Code	Course	Hou wee	k	1	Course Code		Course	Hours/ week			
		L	Т	Р	С			L	T	Р	С
Semester	– I (Courses: 8)	Duration = 15 weeks			Semester	– II (Courses: 7)	Duration = 15 weeks				
MIV501	Cell Biology	1	-	-	1	MIV502	Epidemiology	2	1	-	3
MIV503	Basic Virology	2	1	-	3	MIV504	Molecular Virology and Bioinformatics	2	2	P_	4
MIV505	Biosafety, Biosecurity and Bioethics	2	1	-	3	MIV506	Virological Techniques	2	2	-	4
MIV507	Tissue/Cell culture	1	1	-	2	MIV508	Analytical Tools (Application of GIS and Biostatistics)	1	1	-	2
MIV509	Systematic Virology	4	2	-	6	MIVEL 510.1	Emerging Viral Diseases and Public health response	1	2	-	3
MIV511	Immunology of Viral diseases	2	1	5	3	MIVEL 510.2	One health approach in Virology	1	2	-	3
MIV513	Practical I (Tissue/Cell Culture)	5	-	2	1	MIV512	Practical II (Molecular Virology and Virological techniques)	-	-	6	3
MIV515	Microbiology posting	1	1	-	2	MIV514	Laboratory Rotation-I	-	-	4	2
Total		13	7	2	21	Total		8	8	10	21

L: Lectures; T: Tutorials; P: Practicals; C: Credits



Course Code	Course	Ho we	urs/ ek			Course Course		Ho we	urs/ ek		
		L	Т	Р	С			L	Т	Р	С
Semeste	r – III (Courses: 7)	Duration = 15 weeks			Semeste (Courses			ratio wee			
MIV601	Clinical & Diagnostic Virology -I	2	2	-	4	MIV699	Research Project	-	-	-	18
MIV603	Clinical & Diagnostic Virology -II	2	2	-	4						
MIV605	Viral Vaccines & Antiviral Pharmacotherapy	2	2	-	4		4				
MIV607	Insect vectors of Viral diseases	1	-	2	2		100,				
MIV609	Virology lab design and management	1	-	-	1	118)				
MIV611	Intellectual Property Rights and Patenting	1	-	_ (1						
MIV613	Comprehensive Practical (Clinical and Diagnostic Virology + Laboratory Rotation-II)			8	4						
Total		9	6	10	20	Total		-	-	-	18

L: Lectures; T: Tutorials; P: Practicals; C: Credits



7. DETAILED COURSEWISE INFORMATION

FIRST SEMESTER





CURRICULUM

Manipal Institute of Virology

Name of the Program	MSc Clinical Virology				
Course Title	Cell Biology				
Course Code	MIV501				
Academic Year	2022-2024				
Semester					
Course credits	1				
Course Prerequisite	First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences)				
Course Synopsis	 This module will help To understand basic concept of cell's structure, mechanism and function. To understand the cellular mechanisms through knowledge of cell cycle, cell signalling, and cell differentiation. To know the replication, transcription and translation to understand how they are important to cellular function. To support the student to critically appraise scientific journal, article, review papers, etc. 				
Course Outcomes	CO 1: Outline the basic concepts of eukaryotic and prokaryotic cell structure, mechanism, and function. (C2) CO 2: Understand the cellular biomolecules. (C2) CO 3: Illustrate regulatory pathways in cell cycle, cell signalling and cell differentiation. (C2) CO 4: Illustrate gene organization and chromosomal structure. (C2) CO 5: Outline genetic code on the basis of its principle. (C2) CO 6: Illustrate DNA replication in eukaryotic and prokaryotic cells. (C2) CO 7: Explain RNA synthesis and processing in				



CO 8: Explain translation and post translational modifications of protein synthesis. (C2) **Mapping of COs to POs** РО COs 12 13 7 8 10 11 14 CO 1 ✓ ✓ ✓ CO 2 CO 3 CO 4 ✓ CO 5 ✓ ✓ CO 6 CO 7 ✓ ✓ CO 8

	LEARNING STRATEGY	CONTACT HOURS	SLT
	Lecture	10	30
	Seminar	3	9
	Small Group Discussion (SGD)	1	3
	Self-directed learning (SDL)	1	3
Learning Strategies, Contact Hours and Student Learning Time (SLT)	Problem Based Learning (PBL)	-	-
	Case Based Learning (CBL)	-	-
	Clinic	-	-
	Practical	-	-
	Revision	-	-
	Assessment	1	-
	TOTAL	16	45
	FORMATIVE	SUMMATIVE	
Assessment Methods	Assignment	Mid semeste	r exam
assessificiti Metilous	Student presentation		
	Group discussion		

Mapping of assessment with COs								
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5	CO 6	CO 7	CO 8
Assignments	✓						✓	
Student presentations	✓	✓	✓	✓		✓		
Group discussion		✓		✓	✓			✓
Mid-semester examination	✓	✓	✓	✓		✓		
End-semester examination	×	×	×	×	×	×	*	×
Practical examination	×	×	×	×	×	×	×	×



Feedback Methods	Student feedback on Course and Course master.			
	Cell biology - Gerald Karp			
	The cell – A molecular approach - Cooper			
Reference Materials	Biochemistry - Jeremy M. Berg			
Reference Materials	Molecular biology of the cell - Bruce Alberts			
	International Review of Cell and Molecular Biology -			
	Lorenzo Galluzzi			

Course le	earning outcomes		
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)
Unit 1	Cell type, structure and organelles	 Outline cellular biology. (C2) Recall the history of cell, based on microscopic studies. (C1) Explain the fundamental aspects of cell. (C2) Illustrate the structure and functions of cell. (C2) Identify the key difference between eukaryotic and prokaryotic cells. (C1) Outline the mechanisms of cell organelles contributing to cell function. (C2) Explain the fundamental aspects of the cell organelles and its morphology and mechanism in cell. (C2) 	2/2 = 4
Unit 2	Macromolecules	 Explain the role and type of macromolecules in cell. (C2) Illustrate the structure and functions of macromolecules in cell. (C2) 	1
Unit 3	Cell signalling	 Outline cell signalling. (C2) Classify signalling pathways in cells. (C2) 	1
Unit 4	Cell cycle and Regulation of cell cycle	 Define cell cycle. (C1) Outline the different phases of cell cycle. (C2) 	2/1 = 3



	T		
		 Summarize the fundamental aspects of mitosis and meiosis in cell division. (C2) Introduction to cell cycle. (C1) Illustrate the regulatory pathways in cell cycle. (C2) Explain the role of cyclindependent kinases in cell cycle. (C2) 	
Unit 5	Cell differentiation	 Outline eukaryotic cell differentiation. (C2) Identify the mammalian cell types. (C3) Explain mechanism of cell differentiation. (C2) Summarize epigenetic control over stem cell differentiation. (C2) 	1
Unit 6	Organization of genes and chromosomes	 Explain gene organization in chromosomes. (C2) Illustrate the eukaryotic chromosome structure and function. (C2) Illustrate karyotyping in cytogenetics. (C2) Describe chromosomal aberrations. (C1) 	1
Unit 7	DNA Replication in eukaryotic and prokaryotic cells	 Outline DNA replication. (C2) Explain the different stages of DNA replication in eukaryotes and prokaryotes. (C2) Summarize the role of different enzymatic factors in replication. (C2) 	1
Unit 8	RNA synthesis and processing	 Classify the types of RNA in cell. (C1) Explain the transcription mechanism in eukaryotes and prokaryotes. (C2) Explain the post transcriptional mRNA processing. (C2) 	1



Outline genetic code. (C2) Unit 9 Genetic code • Explain the hypothesis behind 1			T
genetic code. (C2)	Unit 9	Genetic code	• Explain the hypothesis behind 1
Unit 10 Classify proteins involved in cell machinery. (C1) Classify the ribosome machineries in translation. (C2) Explain the different stages of eukaryotic and prokaryotic protein translation. (C2) Summarize the post translational modifications and its maturation in	Unit 10	post translational	 Classify proteins involved in cell machinery. (C1) Classify the ribosome machineries in translation. (C2) Explain the different stages of eukaryotic and prokaryotic protein translation. (C2) Summarize the post translational





CURRICULUM

Name of the Program	MSc Clinical Virology
Course Title	Basic Virology
Course Code	MIV503
Academic Year	2022-2024
Semester	
Course credits	3
Course Prerequisite	First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences)
Course Synopsis	 This module will help To develop an overall idea about the virus evolution, structure, and taxonomy and their interaction with the host. To develop operational and theoretical skills in different microscopic techniques including Electron Microscopy. To critically appraise scientific journal, article, review papers, etc.
Course Outcomes	co 1: Describe the basic concepts of viruses and evolution of viruses. (C4, P1, A2) co 2: Classify viruses based on current classification system. (C2, P1) co 3: Describe the structure of viruses and genome organization. (C2, P1) co 4: Illustrate mathematical modelling of viruses. (C4, P2, A2) co 5: Explain the replication strategies of viruses. (C3, P1) co 6: Understand the principle, use, and application of various types of microscopy in virology. (C4, P3, A3) co 7: Conceptualize the need of electron microscopy in virology. (C3, P1)
Mapping of COs to POs	
IMAPPING OF COS TO FOS	



	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО		РО	РО	PO	РО
COs	1	2	3	4	5	6	7	8	9	10	1	11	12	13	14
CO 1	✓											✓			
СО	√											✓			
2	•														
CO	✓											✓			
СО	√		√						✓			√			
4	-		·												
CO 5	✓		✓									✓			
CO 6	✓				✓	✓						✓			
CO 7	✓				✓	✓						✓			
Learn	ing St	rateg	ies, C	ontac	t Hou	rs	LEAR	NING	ì		COI	NTA	СТ	SLT	
and S	tuder	nt Lea	rning	Time	(SLT)		STRATEGY				но	UR	1.		
							Lecti	ıre			30			90	
							Seminar				8			24	
							Smal	l Gro	Jb dr		3			9	
								ussion)					
							Self-	direct	ed		2			6	
							learn	ing (S	DL)						
							Problem Based				2			6	
							Learning (PBL)								
								Base			-			-	
								ning (CBL)						
							Clinic				-			-	
							Pract				-			-	
18							Revis				-			-	
								ssmei	nt		3			-	
			_	11,			TOTA				48	 		135	
		1						MATI						1ATIVE	
Assessment Methods								nmer						mester	
			•					ent pr			n		End se	mester	exam
		12,					Grou	ıp disc	cussio	n					



Mapping of assessment with CO)s						
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5	CO 6	CO 7
Assignments	✓		✓	✓			
Student presentations					✓	✓	✓
Group discussion	✓		✓	✓			✓
Mid-semester examination	✓	✓	✓	✓			
End-semester examination	✓	✓	✓	✓	✓	✓	✓
Practical examination	×	×	×	×	×	×	×

Feedback Methods	Student feedback on Course and Course master.
Main Reference	 Fields Virology, Knipe David M Principles of Virology Vol 1: Molecular Biology, Flint S J; Others
	 Topley and Wilson's Microbiology and Microbial Infections: Virology Vol 1&2, Mahy Brian W J; Meulen Volker
	Color Atlas of Virology, Versteeg J.
	Journal of Medical VirologyVirology journal

Course le	earning outcomes		
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)
Unit 1	Introduction, History and evolution of "Virology"	 Discuss the history and evolution of virology. (C2, P1, A1) Describe important landmarks in the study of viruses. (C2, P1, A1) Explain the basic concepts and importance of viruses through time. (C4, P1, A2) Introduction to modern virology and its principles. (C4, P2, A2) 	4
Unit 2	Virus structure (general) Virus Replication (general)	 Explain the fundamental aspects of virology – morphology, classification and replication. (C6, P4, A3) 	6/5 = 11



	T		1
		 Describe the general properties of viruses. (C2, P1) Discuss the structure-capsid symmetry and virus architecture of viral particles 	
		 and genome characteristics of the DNA and RNA genome. (C6, P5, A4) Explain the life cycle of virus. (C4, P3, A2) Illustrate/demonstrate handmade virus models for better understanding. (C6, P5, A4) Elucidate the mechanisms viruses use to replicate in their 	
		hosts. (C4, P5, A3)	
Unit 3	Taxonomy and Classification of Viruses	 Explain the classification and taxonomy of viruses. (C4) Explain the Baltimore classification for viruses. (C2) Describe the ICTV classification. (C2) 	3/1 = 4
Unit 4	Bacteriophage- Structure, Replication etc.	 Introduction to bacteriophage. (C2, P1) Describe the distinguishing characteristics of bacteriophage. (C2, P1, A1) Explain about the host specificity and host range of bacteriophage. (C2) Identify modes of infection and phage-host interactions. (C4, P1) Describe the strategies of bacteriophage replication. (C2) Describe the phage life cycles lytic and lysogenic cycles of bacteriophages. (C2) 	4/3 = 7



		 Diagnostic and therapeutic application of bacteriophages. (C3, P1, A2)
Unit 5	Microscopy Light Microscope(y) Fluorescence Microscope(y) Phase Contrast Microscope(y) Inverted Microscope Confocal Microscope (y)	 Discuss the history and introduction. (C1) Describe microscope basics, including parts of light microscope. (C2) Discuss the types and functions of different types of microscopes / Explain the general principles, properties, working and distinguishing features of different types of microscopes. (C3, P1) Understand, set-up and handle microscopes. (C3, P2, A3) Identify the key differences between bright field and dark field microscopy. (C2) Outline different methods used for microscopic staining procedures (histochemical techniques). (C3, P1, A2) Explain the different types of stains used. (C2) Discuss microscope maintenance and best practices for its proper care. (C2) Explain the applications of different types of microscopes. (C3, P1) Explain the advantages and limitations of different types
Unit 6	Electron	of microscopes. (C2) • Understanding the working 4/1 = 5
J.iic 0	Microscope(y) - TEM - SEM	principles of electron microscopy. (C3, P2, A2)



	ı		Г
	- Other variants of EM	 Outline the differences between light microscope and electron microscope. (C4, P1) Describe the theory and applications of electron microscopy. (C4, P2) Explain the principles of operation and basic instrumentation of TEM/SEM. (C3, P3, A2) Demonstrate the theoretical knowledge, working principle, care and use of SEM/TEM. (C4, P2) Specimen preparation for EM – Liquid, tissue and other types of specimen. (C3, P3, A2) Apply and interpret the experimental data. (C4, P1) Understanding other types variants of EM and recent developments. (C3, P1) Development of an EM labfactors for consideration. (C3, P1) 	
Unit 7	EM and its	 Role of EM in virology. (C3, P1) 	3
Jille 7	Applications in Virology	 Critically analyse the use of electron microscopy in virology. (C4, P1) 	3





CURRICULUM

Name of the Program	MSc Clinical Virology
Course Title	Biosafety, Biosecurity and Bioethics
Course Code	MIV505
Academic Year	2022-2024
Semester	
Course credits	3
Course Prerequisite	First-class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences)
Course Synopsis	 This module will help To understand the concepts of biosecurity and biosafety. To provide fundamental knowledge of principles of sterilization, disinfection, decontamination, and biomedical waste management. To understand the importance of ethical principles and practices in scientific research. To provide detailed information about the breeding and maintenance of laboratory animals and genetically modified animals.
Course Outcomes	CO 1: Explain the concept of biosafety and biosecurity. (C2) CO 2: Practice the various methods of sterilization, disinfection, and decontamination. (C3, P2) CO 3: Illustrate safe transportation of infectious materials. (C3, P1) CO 4: Classify biomedical waste. (C2) CO 5: Employ the protocols of biomedical waste management. (C3, P1) CO 6: Develop skills to prevent, mitigate and control laboratory accidents. (C3, P2) CO 7: Explain ethical principles and practices in scientific research. (C2) CO 8: Outline the ethics in animal and human research. (C2)



CO 9: Explain the Central animal research facility available in the University. (C2)

CO 10: Illustrate use of transgenic animals in research. (C2)

CO 11: Develop experimental protocols and project proposals. (C3)

CO 12: Explain application of database and research metrics. (C2)

Mapp	oing o	f COs	to PC)s										
	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	PO
COs	1	2	3	4	5	6	7	8	9	10	11	12	13	14
CO 1	✓		✓					✓	✓		✓			
CO 2	✓		✓			✓					√			
CO 3	✓	✓				✓					✓			
CO 4	✓	✓	✓	✓		✓					✓			
CO 5	✓	✓	✓								√	4 1		
CO 6	✓		✓	✓		✓					V			
CO 7	√										Y	√		✓
CO 8	√										√	✓		
CO 9	√					✓		✓			✓	√		
CO 10	✓					✓		✓	1		✓	✓		
CO 11	✓	✓	✓		✓	V	~	Y	✓		√	✓		
CO 12	√		✓						✓	√	✓	✓		

Learning Strategies, Contact Hours and Student Learning Time (SLT)

LEARNING STRATEGY	CONTACT	SLT
	HOUR	
Lecture	30	90
Seminar	8	24
Small Group	2	6
Discussion (SGD)		
Self-directed learning	3	9
(SDL)		
Problem Based	2	6
Learning (PBL)		
Case Based Learning	-	_
(CBL)		
Clinic	-	_
Practical	-	-
Revision	-	-
Assessment	3	-
TOTAL	48	135



	FORMATIVE	SUMMATIVE	
Assessment Methods	Assignment	Mid semester exam	
	Student presentation	End semester exam	
	Group discussion		

Mapping of assessmen	Mapping of assessment with COs											
	СО	СО	СО	СО	СО	СО	СО	СО	СО	СО	СО	СО
Nature of assessment	1	2	3	4	5	6	7	8	9	10	11	12
Assignments					✓				✓	✓		
Student presentations		✓										√
Group discussion	✓	✓	✓	✓				✓			1	
Mid-semester examination	✓	✓	✓				✓	✓	✓	V	✓	✓
End-semester examination	✓	✓	✓	✓	✓	✓	✓	V	~	✓	✓	✓
Practical examination	×	×	×	×	×	×	×	×	×	×	×	×

Feedback Methods	Student feedback on Course and Course master.
Feedback Methods Main Reference	 Student feedback on Course and Course master. Laboratory Biosafety Manual- WHO Biosafety in Microbiological and Biomedical Laboratories-CDC NIH Guidelines for Biosafety Laboratory Competency-MMWR CDC Regulations and Guidelines on Biosafety of
MANPALI	 Recombinant DNA Research & Biocontainment (DBT, India, 2017) Proposed Framework for the Oversight of Dual Use Life Sciences Research: Strategies for Minimizing the Potential Misuse of Research Information (2007) National Academy of Sciences. On Being a Scientist: A Guide to Responsible Conduct in Research: Third Edition, 2009
	 Research Ethics Training Curriculum, Second Edition by Roberto Rivera and David Borasky ICMR Ethical guidelines for Biomedical Research on Human Participants Fundamentals of Experimental Pharmacology. 3rd ed by M. N. Ghosh



Course le	arning outcomes		
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)
Unit 1	Concept of Biosafety and Biosecurity; Principles of Sterilization and Disinfection	 Discuss biosafety and biosecurity in a laboratory. (C2) Describe the different methods of sterilization. (C2, P1) Distinguish between sterilization and disinfection. (C4) Explain the procedures for decontamination of solid and liquid wastes. (C2, P1) Explain disinfectants and test for disinfectants. (C2) 	4/4 = 8
Unit 2	Biosafety levels	 Describe biosafety level. (C2) Classify risk groups with suitable examples. (C2) 	2/2 = 4
Unit 3	Biosafety cabinets	 Illustrate biosafety cabinets. (C3, P1) Explain HEPA filters and their working principle. (C2) 	2/1 = 3
Unit 4	Biomedical waste and its management	 Explain the need for biomedical waste management system in clinical laboratories. (C6, P1) 	2/1 = 3
Unit 5	Laboratory Containment	 Explain laboratory containment at different biosafety levels. (C3, P1) 	2/2 = 4
Unit 6	Safe Transportation of infectious materials	Describe safe transportation of infectious materials. (C2, P1)	2/1 = 3
Unit 7	Laboratory accidents and its prevention, mitigation, and control	 Describe the use of Personnel Protective equipment. (C2, P1) Develop skills to prevent, mitigate and control laboratory accidents. (C3, P2) 	2/1 = 3
Unit 8	Dual research of concern (DURC)	 Define, outline, and identify the DURC. (C2) Evaluate life sciences research for dual use. (C2) 	2



	T		
Unit 9	Research Ethics	 Assess the risk under potentially dual research of concern and plan risk management. (C3) Model institutional reviews and develop communication plans. (C3, P1) Develop a code of conduct for Dual Research of concern. (C2) Explain the Importance of 	3/1 = 4
Omt 9	Principles of Research Ethics Scientific misconduct Case studies: Violation of scientific standards	 Explain the importance of Research Ethics. (C1) Infer different types of research misconduct. (C2) Demonstrate how scientific misconduct can affect various stakeholders of research. (C2) 	5/1-4
Unit 10	Institutional ethics committees for Animal and Human trials • Human Subject Research Care, breeding and experimentation on Laboratory animals	 Explain the ethics in animal Research. (C1) Explain the ethical principles in research involving humans. (C1) Explain the international declarations on human rights and Human subject Research. (C1) 	3
Unit 11	Visit to the Central Animal House Facility	 Illustrate awareness of the animal research facilities. (C2) Explain key components of the Central Animal House and guidelines followed. (C1) 	2
Unit 12	Genetically modified animals Transgenic animals Cloning Guidelines for the use of genetically modified animals	 Show the theory behind generating Transgenic animals. (C2) Outline different types of vectors and their use in cloning. (C2) Explain the guidelines followed for the use of genetically modified animals. (C1) 	2
Unit 13	Research methods and Protocol development	 Outline the different research methods. (C2) Interpret the basic components of protocol. (C2) 	2



	•	Develop experimental protocols	
		and project proposals. (C3)	
Unit 14 Publication	n Ethics •		2
	•	(C1) Apply plagiarism software like	
		Turnitin. (C3)	
	•	Outline Indexing Database:	
		Scopus. (C2)	





Manipal Institute of Virology

CO 5

Learning Strategies, Contact Hours and

Student Learning Time (SLT)

CURRICULUM

Course Title Course Code MIV507 Academic Year 2022-2024 Semester I No. of credits 2 Course Prerequisite First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences) Course Synopsis This module will help 1. To understand the basic requirements for a cell culture lab. 2. To provide fundamental knowledge of various types of cell lines used for the propagation of viruses. Course Outcomes Course Outcomes Course Code MIV507 Line Substitute Course Course Title Line Substitute Course Course Code Course Course Code Course Course Code Course Course Code Co
Academic Year Semester I No. of credits Course Prerequisite First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences) This module will help 1. To understand the basic requirements for a cell culture lab. 2. To provide fundamental knowledge of various types of cell lines used for the propagation of viruses. Course Outcomes Course Outcomes Course Outcomes
Semester I
No. of credits Course Prerequisite First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences) This module will help 1. To understand the basic requirements for a cell culture lab. 2. To provide fundamental knowledge of various types of cell lines used for the propagation of viruses. Course Outcomes Course Outcomes Course Outcomes Course Outcomes
Course Prerequisite First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences) This module will help 1. To understand the basic requirements for a cell culture lab. 2. To provide fundamental knowledge of various types of cell lines used for the propagation of viruses. Course Outcomes Course Outcomes Course Outcomes
Sciences/Health Sciences) This module will help 1. To understand the basic requirements for a cell culture lab. 2. To provide fundamental knowledge of various types of cell lines used for the propagation of viruses. Course Outcomes Course Outcomes Course Synopsis To provide fundamental knowledge of various types of cell lines used for the propagation of viruses.
To understand the basic requirements for a cell culture lab. To provide fundamental knowledge of various types of cell lines used for the propagation of viruses. Course Outcomes Course Outcomes Course Outcomes
for a cell culture lab. 2. To provide fundamental knowledge of various types of cell lines used for the propagation of viruses. Course Outcomes CO 1: Describe the basic requirements for a
2. To provide fundamental knowledge of various types of cell lines used for the propagation of viruses. Course Outcomes CO 1: Describe the basic requirements for a
various types of cell lines used for the propagation of viruses. Course Outcomes CO 1: Describe the basic requirements for a
propagation of viruses. Course Outcomes CO 1: Describe the basic requirements for a
Course Outcomes CO 1: Describe the basic requirements for a
L coll culture lab (CALAD)
cell culture lab. (C4, A2) CO 2: Identify and differentiate cell culture
methods and types of cell lines. (C4, P3)
CO 3: Demonstrate the techniques of sub
culturing, media preparation, cell counting,
and cell preservation. (C3, P3)
CO 4: Explain the application of cell culture in
virology. (C3, A2)
CO 5: Understand the quality control of cell
lines. (C2, P3)
Mapping of COs to POs
PO
CO 1 V V V V V
CO 2
CO 3
CO 4 V V V



LEARNING

STRATEGY

Lecture

CONTACT

HOUR

15

SLT

45

	Seminar 7 21
	Small Group 2 6
	Discussion
	(SGD)
	Self-directed 5 15
	learning (SDL)
	Problem 1 3
	Based
	Learning (PBL)
	Case Based -
	Learning (CBL)
	Clinic -
	Practical
	Revision -
	Assessment 1
	TOTAL 31 90
Assessment Methods	FORMATIVE SUMMATIVE
	Assignment Mid ser ester exam
	Student presentation
	Group discussion

Mapping of assessment with COs						
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5	
Assignments	✓					
Student presentations		✓	✓		✓	
Group discussion				✓		
Mid-semester examination	✓	✓	✓			
End-semester examination	×	×	×	×	×	
Practical examination	×	×	×	×	×	

Feedback Methods	Student feedback on Course and Course master.
Main Reference	 Culture of Animal Cells by R. Ian Freshney Animal Cell Culture by John R. W. Masters Cell and Tissue Culture by Alan Doyle and J. Bryan Griffiths Fields virology



Course le	arning outcomes		
Content	Topics	Learning Outcomes	Hours (Lecture/Tutorials)
Unit 1	Introduction to cell culture, Basic equipment and facilities	 Discuss the terms cell culture, cell line and cell strain. (C2, P1, A1) Discuss the different types of cell culture techniques and classification of mammalian cell lines. (C2, P1, A2) Explain the advantages and limitations of cell culture. (C2) Discuss the biosafety, aseptic techniques in a tissue culture laboratory. (C4, A2) Describe the layout and equipment required for a tissue culture laboratory. (C4, A2) Describe the morphology of cells in culture. (C4, P3, A2) 	4/4 = 8
Unit 2	Cell culture media and Supplements	 Describe the growth requirements for eukaryotic cells, including the culture environment, media and supplements and their preparation. (C3, P3, A2) 	1/3 = 4
Unit 3	Cell counting and sub culturing of cell lines	 Explain the principle and procedure for cell counting (C3, P3, A2) Explain the life span, growth cycle and sub culturing of cell lines and its importance. (C3, P3, A2) 	2/2 = 4
Unit 4	Characterization of cell culture	Identify the different types of cell culture and characterization mainly based on morphology. (C4, P3, A2)	1/2 = 3
Unit 5	Preservation of cell cultures	 Discuss the principle and methods of preservation of cell lines, thawing and revival of frozen cells. (C2, P3, A2) 	1/2 = 3
Unit 6	National / International cell culture collections	Describe the various National and International Cell culture collections and their role. (C1)	1



	Genetic modification of cell lines	Describe the method and principle of modified cell cultures such as shell vial culture, mixed cell culture, genetically engineered cell lines etc. with examples and its applications. (C3, A2)	1
Unit 8	Quality control of cell lines	 Discuss the measures taken to preserve the quality of reagents and cells. (C2) Identify the potential sources, types of contamination and control of contamination. (C4, P3, A2) 	2
Unit 9	Cell culture in Virology	Describe the application of cell culture in Virology. (C4, A2)	1/2 = 3
Unit 10	Large scale production of cell cultures	Discuss about the methods of large- scale culture of cell lines, its production and applications. (C1)	1





Manipal Institute of Virology

CURRICULUM

Name of the Program	MSc Clinical Virology			
Course Title	Systematic Virology			
Course Code	MIV509			
Academic Year	2022-2024			
Semester	1			
Course credits	6			
Course Prerequisite	First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences)			
Course Synopsis	 The course will offer in-depth knowledge about the taxonomy, structure, genome, antigens, replication, animal/cell susceptibility. The course will provide an overview of epidemiology, clinical diseases, pathogenesis, lab diagnosis and prophylaxis. 			
Course Outcomes	CO 1: Explain the difference in structures between the enveloped and non-enveloped viruses. (C2) CO 2: Recall various families of DNA, RNA and Retroviruses. (C1) CO 3: Explain the replication strategies and interaction of DNA viruses, RNA viruses and prions with the host. (C2) CO 4: Illustrate the role of different viral proteins in attachment, fusion, uncoating, replication, assembly, and release. (C2)			
Mapping of COs to POs				
PO PO PO PO PO PO PO COs 1 2 3 4 5 6	PO PO<			
CO 1	, 6 5 16 11 12 13 14 V			
CO v				
2 V				
CO 4 1	✓ ✓ ✓			
Learning Strategies, Contact Hours and Student Learning Time (SLT)	LEARNING CONTACT SLT STRATEGY HOUR			



	Lecture	45	135	
	Seminar	39	117	
	Small Group	3	9	
	Discussion (SGD)			
	Self-directed learning	3	9	
	(SDL)			
	Problem Based	-	-	
	Learning (PBL)			
	Case Based Learning	-	-	
	(CBL)			
	Clinic	1	-	
	Practical	-	_	
	Revision	-	-	
	Assessment	3	-	
	TOTAL	93	270	
Assessment Methods	FORMATIVE	SUI	MMATIVE	
	Assignment	Mid-semes	ter exam	
	Student presentation	End semes	ter exam	
	Group discussion			

Mapping of assessment with COs				
Nature of assessment	CO 1	CO 2	CO 3	CO 4
Assignments	✓			
Student presentations	✓		✓	✓
Group discussion	✓		✓	✓
Mid-semester examination	✓	✓		
End-semester examination	✓	✓	✓	✓
Practical examination	×	×	×	×

Feedback Methods	Student feedback on Course and Course master.	
Main Reference	 Fields Virology, Vol 1 & 2 Principles of Virology, J Flint, Vol 1 & 2 	



Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)
Unit 1	Introduction to DNA and RNA viruses and replication	 Classify DNA and RNA virus families. (C2) Explain the general replication strategies of DNA and RNA viruses. (C2) 	2/1 = 3
Unit 2	Adenoviridae	 Classify the viruses in the Adenoviridae family. (C2) List the diseases caused by viruses in Adenoviridae. (C1) Illustrate the structure of Adenoviruses. (C2) Explain the replication of Adenovirus. (C2) Summarize in detail about the proteins involved in Adenovirus virus replication. (C2) Explain the epidemiology and pathogenesis of Adenoviruses. (C2) Explain the laboratory diagnosis and prophylaxis available for the viruses in Adenoviridae. (C2) 	2/2 = 4
Unit 3	Parvoviridae	 Classify the viruses in Parvoviridae family. (C2) List the diseases caused by viruses in Parvoviridae. (C1) Illustrate the structure of Parvoviruses. (C2) Explain the replication of Primate erythroparvovirus 1. (C2) Summarize in detail about the proteins involved in Primate erythroparvovirus 1 replication. (C2) 	2/1 = 3



		 Explain the epidemiology and pathogenesis of Parvoviruses. (C2) Explain the laboratory diagnosis and prophylaxis available for the viruses in Parvoviridae. (C2)
Unit 4	Poxviridae	 Classify the viruses in Poxviridae family. (C2) List the diseases caused by viruses in Poxviridae. (C1) Illustrate the structure of Poxviruses. (C2) Explain the replication of Poxvirus. (C2) Summarize in detail about the proteins involved in Poxvirus replication. (C2) Explain the epidemiology and pathogenesis of Poxviruses. (C2) Explain the laboratory diagnosis and prophylaxis available for the viruses in Poxviridae. (C2)
Unit 5	Herpesviridae	 Classify the viruses in Herpesviridae family. (C2) List the diseases caused by Herpesviridae. (C1) Illustrate the structure of Herpesviruses. (C2) Explain the replication of Human alpha herpesviruses. (C2) Summarize in detail about the proteins involved in Herpesvirus replication. (C2) Explain the epidemiology and pathogenesis of Herpesviruses. (C2)



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		 Explain the laboratory diagnosis and prophylaxis available for the viruses in Herpesviridae. (C2) Explain the latency of Herpesviruses. (C2)
Unit 6	Papillomaviridae	 Classify the viruses in the Papillomaviridae family. (C2) List the diseases caused by viruses in Papillomaviridae. (C1) Illustrate the structure of Papillomaviruses. (C2) Explain the replication of Human Papillomavirus. (C2) Summarize in detail the proteins involved in Human Papillomavirus replication. (C2) Explain the epidemiology and pathogenesis of Papillomaviruses. (C2) Explain the laboratory diagnosis and prophylaxis available for the viruses in Papillomaviridae. (C2)
Unit 7	Polyomaviridae	 Classify the viruses in the Polyomaviridae family. (C2) List the diseases caused by viruses in Polyomaviridae. (C1) Illustrate the structure of Polyomaviruses. (C2) Explain the replication of Human Polymoviruses. (C2) Summarize in detail the proteins involved in Human Polyomavirus replication. (C2) Explain the epidemiology and pathogenesis of Polyomaviruses. (C2)



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		Explain the laboratory diagnosis and prophylaxis available for the viruses in Palvanaging (C2)
		Polyomaviruses. (C2)
Unit 8	Retroviruses – HIV structure and replication	 Classify the retroviruses and explain the replication strategies of retroviruses. (C2) List the diseases caused by Human Immunodeficiency Virus (HIV). (C1) Illustrate the structure of HIV. (C2) Explain the replication of HIV. (C2) Summarize in detail about the proteins involved in HIV replication and its reverse transcriptase activity. (C2) Explain the epidemiology and pathogenesis of HIV. (C2) Explain the evasion from host immune response by HIV. (C2) Explain the laboratory
		diagnosis and prophylaxis
		available for HIV. (C2)
Unit 9	Hepadnaviridae	 Classify the viruses in the Hepadnaviridae family. List the diseases caused by Hepatitis B virus (HBV). (C1) Illustrate the structure of the HBV. (C2) Explain the replication of HBV. (C2) Summarize in detail about the proteins involved in HBV replication. (C2) Explain the epidemiology and pathogenesis of HBV. (C2) Explain the laboratory diagnosis and prophylaxis available for HBV. (C2)



Unit 10	Flaviviridae	 Classify the viruses in the Flaviviridae family. (C2) List the diseases caused by viruses in Flaviviridae. (C1) Illustrate the structure of Flaviviruses. (C2) Explain the replication of Dengue virus and Hepatitis C virus. (C2) Summarize in detail the proteins involved in Dengue virus and Hepatitis C virus replication. (C2) Explain the epidemiology and pathogenesis of Flaviviridae. (C2) Explain the laboratory diagnosis and prophylaxis available for viruses in Flaviviridae. (C2) 	2/2 = 4
Unit 11	Picornaviridae	 Classify the viruses in Picornaviridae family. (C2) List the diseases caused by viruses in Picornaviridae. (C1) Illustrate the structure of Picornaviruses. (C2) Explain the replication of Enterovirus. (C2) Summarize in detail about the proteins involved in Enterovirus replication. (C2) Explain the epidemiology and pathogenesis of Picornaviridae. (C2) Explain the laboratory diagnosis and prophylaxis available for viruses in Picornaviridae. (C2) 	2/2 = 4
Unit 12	Reoviridae	Illustrate the structure of Reoviruses. (C2)	2/1 = 3



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		 List the diseases caused by viruses in Reoviridae. (C1) Explain the replication of Rotavirus. (C2) Summarize in detail the proteins involved in Rotavirus replication. (C2) Explain the epidemiology and pathogenesis of Reoviridae. (C2) Explain the laboratory diagnosis and prophylaxis available for viruses in Reoviridae. (C2) 	
Unit 13	Filoviridae	 Classify the viruses in the Filoviridae group. (C2) List the diseases caused by viruses in Filoviridae. (C1) Illustrate the structure of Filoviruses. (C2) Explain the replication of the Ebola virus. (C2) Summarize in detail about the proteins involved in Ebola virus replication. (C2) Explain the epidemiology and pathogenesis of Filoviridae. (C2) Explain the laboratory diagnosis and prophylaxis available for viruses in Filoviridae. (C2) 	
Unit 14	Coronaviridae	 Classify the viruses in Coronaviridae family. (C2) List the diseases caused by viruses in Coronaviridae. (C1) Illustrate the structure of Coronaviruses. (C2) Explain the replication of Coronavirus. (C2) 	



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		 Summarize in detail about the proteins involved in Coronavirus replication. (C2) Explain the epidemiology and pathogenesis of Coronaviridae. (C2) Explain the laboratory diagnosis and prophylaxis available for viruses in Coronaviridae. (C2) 	
Unit 15	Paramyxoviridae	 Classify the viruses in Paramyxoviridae family. (C2) List the diseases caused by viruses in Paramyxoviridae. (C1) Illustrate the structure of Mumps virus, Measles virus, Parainfluenza virus and Nipah virus. (C2) Explain the replication of different viruses in the Paramyxoviridae family. (C2) Summarize in detail about the proteins involved in the replication of viruses in Paramyxoviridae family. (C2) Explain the epidemiology and pathogenesis Paramyxoviridae. (C2) Explain the laboratory diagnosis and prophylaxis available for viruses in 	2/2 = 4
Unit 16	Pneumoviridae	Paramyxoviridae. (C2) • Classify the viruses in	2/1 = 3
		 Pneumoviridae family. (C2) List the diseases caused by viruses in Pneumoviridae. (C1) Illustrate the structure of Respiratory Syncytial virus and Metapneumovirus (C2) 	_, _ S



		 Explain the replication of different viruses in the Pneumoviridae family. (C2) Summarize in detail about the proteins involved in the replication of viruses in the Pneumoviridae family. (C2) Explain the epidemiology and pathogenesis Pneumoviridae(C2) Explain the laboratory diagnosis and prophylaxis available for viruses in Pneumoviridae. (C2)
Unit 17	Hantaviridae	 Classify the viruses in Hantaviridae family. (C2) List the diseases caused by viruses in Hantaviridae. (C1) Illustrate the structure of Human Hanta viruses(C2) Explain the replication of different viruses in the Hantaviridae family. (C2) Summarize in detail about the proteins involved in the replication of viruses in the Hantaviridae family. (C2) Explain the epidemiology and pathogenesis Hantaviridae(C2) Explain the laboratory diagnosis and prophylaxis available for viruses in Hantaviridae. (C2)
Unit 18	Nairoviridae	 Classify the viruses in Nairoviridae family. (C2) List the diseases caused by viruses in Nairoviridae. (C1) Illustrate the structure of Nairoviruses. (C2)



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		 Explain the replication of Crimean-Congo hemorrhagic fever orthonairovirus. (C2) Summarize in detail about the proteins involved in Crimean-Congo hemorrhagic fever orthonairovirus replication. (C2) Explain the epidemiology and pathogenesis of Nairoviridae. (C2) Explain the laboratory
		diagnosis and prophylaxis available for viruses in Nairoviridae. (C2)
Unit 18	Orthomyxoviridae	 Classify the viruses in Orthomyxoviridae family. (C2) List the diseases caused by viruses in Orthomyxoviridae. (C1) Illustrate the structure of Orthomyxoviruses. (C2) Explain the replication of Influenza virus. (C2) Summarize in detail about the proteins involved in Influenza virus replication. (C2) Explain the epidemiology and pathogenesis of Orthomyxoviridae. (C2) Explain the laboratory diagnosis and prophylaxis available for viruses in Orthomyxoviridae. (C2)
Unit 19	Rhabdoviridae	 Classify the viruses in Rhabdoviridae family. (C2) List the diseases caused by viruses in Rhabdoviridae. (C1) Illustrate the structure of Rabies lyssavirus. (C2)



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		 Explain the replication of Rabies lyssavirus. (C2) Summarize in detail about the proteins involved in Rabies lyssavirus replication. (C2) Explain the epidemiology and pathogenesis of Rhabdoviridae. (C2) Explain the laboratory diagnosis and prophylaxis available for viruses in Rhabdoviridae. (C2) 	
Unit 20	Astroviridae	 Classify the viruses in Astroviridae family. (C2) List the diseases caused by viruses in Astroviridae. (C1) Illustrate the structure of Mamastrovirus 1. (C2) Explain the replication of Mamastrovirus 1. (C2) Summarize in detail about the proteins involved in Mamastrovirus 1 replication. (C2) Explain the epidemiology and pathogenesis of Astroviridae. (C2) Explain the laboratory diagnosis and prophylaxis available for viruses in Astroviridae. (C2) 	2/1 = 3
Unit 21	Caliciviridae	 Classify the viruses in Caliciviridae family. (C2) List the diseases caused by viruses in Caliciviridae. (C1) Illustrate the structure of Norwalk virus. (C2) Explain the replication of Norwalk virus. (C2) 	2/1 = 3



		Summarize in detail about the
		proteins involved in Norwalk virus replication. (C2) Explain the epidemiology and
		pathogenesis of Caliciviridae. (C2)
		 Explain the laboratory diagnosis and prophylaxis available for viruses in Caliciviridae family. (C2)
Unit 22	Prions and slow viral diseases	 List different viruses causing slow viral infections. (C1) List the diseases caused by Prions and slow viral infections. (C1) Explain the structure and formation of cellular prion protein. (C2) Explain in detail about the change from PrP^C to PrP^{Sc}. Explain the epidemiology and pathogenesis of Prion diseases. (C2)
	اک،	 Explain the laboratory diagnosis and prophylaxis available for Prions. (C1)
Unit 23	Togaviridae	 Classify the viruses in Togaviridae family. (C2) List the diseases caused by viruses in Togaviridae. (C1) Illustrate the structure of Togaviruses. (C2) Explain the replication of Chikungunya virus. (C2) Summarize in detail about the proteins involved in Chikungunya virus replication. (C2) Explain the epidemiology and pathogenesis of Togaviridae. (C2)



		Explain the laboratory diagnosis and prophylaxis available for viruses in Togaviridae. (C2)
Unit 24	Matonaviridae	 Classify the viruses in Matonaviridae family. (C2) List the diseases caused by viruses in Matonaviridae. (C1) Illustrate the structure of Togaviruses. (C2) Explain the replication of the Rubella virus. (C2) Summarize in detail the proteins involved in Rubella virus replication. (C2) Explain the epidemiology and pathogenesis of Matonaviridae. (C2) Explain the laboratory diagnosis and prophylaxis available for viruses in Matonaviridae. (C2)
Unit 25	Arenaviridae	 Classify the viruses in Arenaviridae family. (C2) List the diseases caused by viruses in Arenaviridae. (C1) Illustrate the structure of Arenaviruses. (C2) Explain the replication of Lymphocytic choriomeningitis mammarenavirus. (C2) Summarize in detail about the proteins involved in Lymphocytic choriomeningitis mammarenavirus replication. (C2) Explain the epidemiology and pathogenesis of Arenaviridae. (C2) Explain the laboratory diagnosis and prophylaxis



		available for viruses in	
		Arenaviridae. (C2)	
	D 1: 1	, ,	4/4 2
Unit 26	Deltavirus	 List the diseases caused by Hepatitis delta virus. (C1) Illustrate the structure of Hepatitis delta virus. (C2) Explain the replication of Hepatitis delta virus. (C2) Summarize in detail about the proteins involved in Hepatitis delta virus replication. (C2) Explain the epidemiology and pathogenesis of Hepatitis delta virus. (C2) Explain the laboratory diagnosis and prophylaxis available for Hepatitis Delta 	1/1 = 2
		virus. (C2)	
Unit 27	Hepeviridae	 Classify the viruses in Hepeviridae family. (C2) List the diseases caused by viruses in Hepeviridae. (C1) Illustrate the structure of Orthohepevirus A. (C2) Explain the replication of Orthohepevirus A. (C2) Summarize in detail about the proteins involved in Orthohepevirus A replication. (C2) Explain the epidemiology and pathogenesis of Hepeviridae. (C2) Explain the laboratory diagnosis and prophylaxis available for viruses in Hepeviridae. (C2) 	1/1 = 2





Name of the program	MSc Clinical Virology	
Course Title	Immunology of Viral Diseases	
Course Code	MIV511	
Academic Year	2022-2024	
Semester	1	
Course credits	3	
Course Prerequisite	First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences)	
Course Synopsis	 This module helps the student to understand the basic principles and key concepts of immunology. It gives an overview of cellular and molecular events that control the function of immune system such as immune detection, activation and response to an infection. This course also emphasizes the host-virus interaction and specific immune response upon viral entry. 	
	4. This course imparts understanding of immunopathogenesis of viral diseases.	
Course Outcomes	CO 1: Explain the different cells and organs of immune system. (C1) CO 2: Differentiate cellular and humoral immunity. (C2) CO 3: Describe the process of B-cell and T-cell synthesis, maturation, activation, selection, proliferation and response. (C2) CO 4: Elaborate the activators and suppressors of immune system. (C2) CO 5: Define the specific immune response triggered upon viral infections. (C3) CO 6: Explain cytokines in viral infections. (C2) CO 7: Illustrate complement pathway. (C2)	



CO 8: Outline antibody dependent enhancement. (C2)

CO 9: Analyse recognition of viruses by cellular sensors. (C4)

CO 10: Explain specific innate and adaptive response to viral infections and vaccination. (C5)

CO 11: Outline principle, instrumentation and analytical applications of Flow cytometry. (C2)

Mappir	ng of C	Os to	POs											
COs	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO 1	✓										✓			
CO 2	✓		✓								✓			
CO 3	✓		✓					✓			✓			
CO 4	✓										✓			
CO 5	✓							✓			V			
CO 6	✓		✓								✓			
CO 7	✓										✓			
CO 8	✓									P	✓			
CO 9	✓		✓					✓			✓			
CO 10	✓	_	✓					✓	✓		✓			
CO 11	✓		✓					V			✓			

Learning Strategies, Contact Hours		LEARNING STRATEGY	CONTACT	SLT
and Student Learning Time (SLT)	ď	Y .	HOUR	
	П	Lecture	30	90
	DĪ	Seminar	7	21
		Small Group	5	15
		Discussion (SGD)		
		Self-directed learning	2	6
		(SDL)		
		Problem Based	-	-
		Learning (PBL)		
		Case Based Learning	-	-
		(CBL)		
		Clinic	-	-
		Practical	-	-
		Revision	-	-
		Assessment	3	-
		TOTAL	47	132
Assessment Methods		FORMATIVE	SUN	IMATIVE
		Assignment	Mid semes	ster exam
		Student presentation	End semes	ter exam
		Group discussion		



Mapping of assessment	with CO	Os									
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5	CO 6	CO 7	CO 8	CO 9	CO 10	C O 11
Assignments			✓				✓	✓	✓		
Student presentations		✓								✓	✓
Group discussion					✓			✓	✓		
Mid-semester examination	✓	✓	✓	✓	✓						√
End-semester examination	✓	√	✓	√	✓	✓	✓	√	✓	~	V
Practical examination	×	×	×	×	×	×	×	×	×	×	×

Feedback Methods	Student feedback on Course and Course master.
Main Reference	Kuby's Immunology
	Roitt's Essential Immunology
	Cellular and molecular immunology: Abdul K. Abbas
	Microbiology and Immunology: Subhash Chandra Parija

Course le	arning outcomes		
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)
Unit 1	1. Introduction to immunology	Make the student to understand the following. (C1)	2
		Basic terminologies.History of immunology.Types of immunity.	
MI	2. Immune system – structure and components	Explain the organisation of immune system. (C2) • Cell of the immune	
	(Immunoglobulins, cells, complement etc)	 system. Tissues and organs of immune system. Antigen. Antibody. Antigen-antibody reaction. 	3



		• Complement system,	
		activation, regulation	
		and its effect.	
Unit 2	Immune response	Distinguish and interpret the	
		various immune responses.	
		(C2)	
		Humoral immunity	0.40
ı		B-cell synthesis,	3/2 = 5
		maturation,	
		differentiation and	
		activation.	
		Cell-mediated immunity.	
		T-cell synthesis,	1 111.
		maturation,	-11
		differentiation and	
		activation.	
		Major Histocompatibility	
		Complex.	
		Transfer factor.	
		Immunological	
		tolerance.	
Unit 3	Immunogenetics	Understand how diversity of	
		T-cell receptors and	
		antibodies arises. (C2)	
		Genetic mechanisms	
	. ()	generating T cell	2
	(1)	receptor and antibody	
		diversity.	
		Antibody variable	
	.012	regions, and gene	
		rearrangement.	
		Impact of genetic	
		variation at individual	
		and population level on	
		susceptibility or	
		resistance to diseases.	
Unit 4	1 Uning and a set to the control of		1
Oilit 4	1. Hypersensitivity	Explain Type I—IV hypersensitivity. (C2)	1
	2 June 100 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	Define the	
	2. Immunodeficiency	immunodeficiency and its	1
		types. (C2)	1
		τγρεσ. (C2)	



• Primary	
immunodeficiency.	
Secondary	
immunodeficiency.	
3. Hybridoma Apply acquired theoretical	
technology knowledge to develop mAbs	2
in future. (C3)	
Principle, method and	
applications of	
hybridoma technology.	
Unit 5 Host-virus Develop the hypothesis of	
interactions immune response for any	
viruses. (C3)	5
Cellular receptors and	
virus entry virus	
morphogenesis.	
Host cell damage Cellular	
and viral gene	
expression.	
Unit 6 Recognition of viruses • Compare different	1/1 = 2
by cellular sensors pathogen recognition	
receptors (PRRs). (C2)	
Illustrate signal	
transduction mechanism	
of PRRs. (C2)	
Unit 7 Overview of Cytokines • Outline cytokines and	1/1 = 2
in Viral Infections cytokine nomenclature.	, –
(C2)	
• Explain properties of	
cytokines. (C2)	
Illustrate cytokine-	
mediated effects. (C2)	
• Summarize cytokine	
actions. (C2)	
Illustrate regulation of	
immune response by	
cytokines. (C2)	
Demonstrate signal	
Transduction by cytokine	
receptors. (C2)	



	Г	Γ	T
		Infer cytokines in viral	
		, ,	
Unit 8	Complement pathway	 infections. (C2) Outline complement system. (C2) Illustrate pathways of complement system. (C2) Classical pathway (specific immune system). Alternative (non-specific immune system). Lectin pathway. Explain protective roles for Complement during Viral Infections. (C2) Outline role of complement system in 	2
		enhancement of	
		humoral immunity to	
		viruses. (C2)	
Unit 9	Antibody dependent enhancement	 Explain mechanism of antibody dependent enhancement. (C2) Demonstrate ADEs in different viral infections. (C2) 	2
Unit 10	Immune response to Viral vaccines	 Explain vaccinemediated protection. (C5) Outline main effectors of vaccine response. (C4) Interpret activation of innate to adaptive immunity in response to vaccination. (C4) Demonstrate vaccine antibody response. (C3) 	2/1 = 3



Unit 11	Viral evasion of	Explain mechanisms of	3
	immune response	viral evasion. (C2)	-
		• Explain Virus specific	
		evasion. (C2)	
Unit 12	Specific immune	Explain specific immune	9
	response to viral	response to (C2)	
	diseases	 Coronavirus 	
		 Influenza 	
		• HIV	
		 Hepatitis 	
		• Dengue	
		 Japanese Encephalitis 	
		virus	
		 Chikungunya virus 	
		• Zika	
		Ebola virus	
		• Kyasanur Forest	
		Disease virus	
Unit 13	Flow cytometry	• Outline principle of Flow cytometry. (C2)	1
		 Illustrate 	
		instrumentation. (C2)	
		 List analytical 	
		applications. (C2)	





Name of the Program							MSc Clinical Virology								
Course Title						Practical I (Tissue/Cell Culture)									
Course Code					MIV513										
Academic Year				202	2-2024	4									
Semester No. of credits					I										
					1										
Cours	se Prer	equisi	ite				Firs	t class	/CGP/	4 6.5	at U	G leve	el (BS	c Life	
							Scie	nces/I	Health	Scien	ces)				
Course Synopsis					 This module will help to understand the basic requirements for a cell culture lab. The course will provide fundamental knowledge of various types of cell lines used for the propagation of viruses. 										
Course Outcomes				CO 1: Identify and differentiate cell culture methods and types of cell lines. (P3) CO 2: Demonstrate the techniques of sub culturing, media preparation, cell counting, and cell preservation. (P3) CO 3: Understand the quality control of cell lines. (C2, P3)											
	ping o							200		РО	РО	РО	РО	РО	
COs CO 1	PO1 ✓	PO2	PO3	PO4	PO5	PO6 ✓	PO7	PO8	PO9 ✓	10	11	12	13	14	
CO 2	~		✓			✓					✓		✓		
2	со										✓				
	✓		<u></u>		Learning Strategies, Contact Hours and Student Learning Time (SLT)					LEARNING STRATEGY			SLT		
co 3 Learn	_	_			Hour	s and						ITACT DUR	SLT		
CO 3 Learn	_	_			Hour	s and	l	STR cture					SLT -		
CO 3 Learn	_	_			Hour	s and	Se	STR. cture minar							
co 3 Learn	_	_			Hour	s and	Se	STR. cture minar	ATEG)	roup					



	Problem Based	-	-	
	Learning (PBL)			
	Case Based	-	-	
	Learning (CBL)			
	Clinic	-	-	
	Practical	30	90	
	Revision	-	-	
	Assessment	2	-	
	TOTAL	32	90	
Assessment Methods	FORMATIVE	SUMN	MATIVE	
	Assessment of	-		
	Record			
	books			
	Internal Assessmen	t		

Mapping of assessment with COs			
Nature of assessment	CO 1	CO 2	CO 3
Record books, technical skills, conduct in lab	✓	✓	✓
Internal assessment	✓	✓	✓
Assignments	×	×	×
Student presentations	×	×	×
Group discussion	×	×	×
Mid semester exam	×	×	×
End semester exam	×	×	×

Feedback Methods	Student feedback on Course and Course master
Main Reference	Culture of Animal Cells by R. Ian Freshney
10/	Animal Cell Culture by John R. W. Masters
	 Cell and Tissue Culture by Alan Doyle and J. Bryan Griffiths Fields virology



Course learning outcomes						
Topics	Learning Outcomes	Hours (Practical)				
Cell culture media and Supplements	 Describe the growth requirements for eukaryotic cells, including the culture environment, media and supplements and their preparation. (P3) 	7				
Cell counting and sub culturing of cell lines	 Explain the principle and procedure for cell counting. (P3) Explain the life span, growth cycle and sub culturing of cell lines and its importance. (P3) 	7				
Characterization of cell culture	Identify the different types of cell culture and characterization mainly based on morphology. (P3)	7				
Preservation of cell cultures	 Discuss the principle and methods of preservation of cell lines, thawing and revival of frozen cells. (P3) 	7				
Cell culture in Virology	 Describe the application of cell culture in Virology. (C4, A2) 	2				





Manipal Institute of Virology

CURRICULUM

Name of the Program						MSc Clinical Virology								
Course Title N								obiolo	gy Po	sting				
Course	e Code						MIV	515				-		
Acade	mic Ye	ar					2022	-2024						
Semes	ster						I							
No. of	credit	S					2							
Course	e Prere	equisit	е									leve	l (BSc	Life
Course Synopsis						First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences) This is a mandatory learning course (MLC), which will be designed and implemented by the Department of Microbiology, KMC, Manipal. It is credited but not graded. Transcript will show S/NS (Satisfactory/Not satisfactory) based on the participation and comprehensive evaluation. The training module enables students to comprehend the fundamental microbiology practices and the diverse facets of infectious disease diagnosis. Core areas of medical microbiology namely immunology, serology, ICTC/HIV surveillance, aerobic bacteriology, anaerobic bacteriology, mycobacteriology, mycology, parasitology, molecular biology of infectious diseases, and microbial surveillance in hospital and community settings are								
Course	e Outc	omes						-				rainin	g in	
						diagnostic microbiology. CO2: To highlight rationale of testing and								
							interpretation.							
								•		ands (on exn	erienc	e in h	asic
							CO3: To impart hands on experience in basi microbiology techniques.					20.0		
1	Mapping of COs to POs							<u>.,</u>	73.					
Mapp	oing of									РО	РО	РО	РО	
								DOG	PO9	10	1 4 4			РО
COs	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8		10	11	12	13	PO 14
COs	PO1			PO4	PO5	✓	PO7	PU8	√	10	✓	12		
COs	PO1		PO3 ✓	PO4	PO5		PO7	PU8		10		12	13 ✓	



LEARNING STRATEGY	CONTACT HOUR	SLT
Lecture	15	45
Seminar	-	-
Small Group Discussion (SGD)	-	-
Self-directed	-	-
Problem Based	-	-
Case Based Learning (CBL)	-	- 1
Clinic	-	-
Practical	15	45
Revision	-	7
Assessment		-
TOTAL	30	90
O.		
	STRATEGY Lecture Seminar Small Group Discussion (SGD) Self-directed learning (SDL) Problem Based Learning (PBL) Case Based Learning (CBL) Clinic Practical Revision Assessment	STRATEGY Lecture 15 Seminar - Small Group Discussion (SGD) Self-directed learning (SDL) Problem Based Learning (PBL) Case Based Learning (CBL) Clinic - Practical Revision - Assessment



SECOND SEMESTER





Name of the Program	MSc Clinical Virology
Course Title	Epidemiology
Course Code	MIV502
Academic Year	2022-2024
Semester	II
Course credits	3
Course Prerequisite	First class/CGPA 6.5 at UG level (BSc Life
course i rerequisite	Sciences/Health Sciences)
Course Synopsis	 This module introduces the principles in epidemiology and public health surveillance. Basic concepts in infectious disease epidemiology. Epidemiological study designs and its applications. Various steps of investigating an outbreak and management. Case-study based training to deepen the knowledge in applied epidemiology.
Course Outcomes	CO 1: Define epidemiology and its applications in public health. (C1) CO 2: Outline basic terminologies used in disease occurrence and basic concepts in infectious disease epidemiology. (C2) CO 3: Elaborate epidemiological study designs. (C6) CO 4: Outline basic concepts in public health surveillance and identify the attributes and limitations of good surveillance mechanism. (C2) CO 5: Outline fundamental principles and steps involved in investigating an outbreak. (C2) CO 6: Utilize EpiInfo software to gather, analyse, and present data. (C3) CO 7: Evaluate and discuss case studies in applied epidemiology. (C5)



Марр	ing of	COs to	POs						,			,		
COs	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO 1	√ · · · · · · · · · · · · · · · · · · ·	102	√ ·	1.04	1.03	1.00	107	100	103	10	√	12	13	14
CO 2	√		√					√			√			
CO 3	✓	✓									✓			
CO 4	✓	✓	✓	✓			✓		✓		✓	✓		✓
CO 5	✓	✓	✓	✓			✓			✓	✓	✓		✓
CO 6	✓		✓	✓	✓					✓	✓			
CO 7	✓		✓						✓		✓			
				LEA	ARNIN	G STRA	ATEGY		NTAC OURS	1	SL	Г		
						Lec	ture				20	1	60)
						Ser	Seminar						15	
							Small Group Discussion (SGD)						9	
						Sel	Self-directed learning (SDL)						9	
	_	itegies Learni				Pro		4		12	!			
						Cas (CB	10			30				
						 	Clinic				-			
					Pra	Practical				-				
				Rev	vision			-			-			
				Ass	essme	nt			3		-			
						TO	TAL				48		13	5
						FO	RMAT	IVE		S	UMN	1ATI\	/E	
SSASS	ment l	Metho	ds				signme			N	∕lid se	mest	er ex	am
133633	ciic i	1100	us				ident p			E	nd se	mest	er ex	am
						Gro	oup dis	cussio	n					

Mapping of assessment with COs	5						
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5	CO 6	CO 7
Assignments						✓	✓
Student presentations		✓	✓	✓	✓		
Group discussion							✓
Mid-semester examination	✓	✓	✓	✓			
End-semester examination	✓	✓	✓	✓	✓	✓	✓
Practical examination	×	×	×	×	×	×	×



 ✓ Principles of Epidemiology in Public Health Practice (3rd edition) ✓ Textbook of Preventive & Social Medicine (by K Park) ✓ Epidemiology (by Leon Gordis) ✓ National Health Programs of India National Policies and Legislations Related to Health (by J. Kishore). ✓ Chapter 53: Public Health Surveillance: A Tool for Targeting and Monitoring Interventions. Available at: https://www.ncbi.nlm.nih.gov/books/NBK11770/pdf/Book shelf_NBK11770.pdf ✓ The CDC Field Epidemiology Manual (by Sonja A. Rasmussen and Richard A. Goodman) ✓ Control of Communicable Diseases Manual (20th Edition) ✓ Modern Epidemiology (3rd edition) (by Kenneth J. 	Feedback Methods	Student feedback on Course and Course master.
Dothman)		 ✓ Principles of Epidemiology in Public Health Practice (3rd edition) ✓ Textbook of Preventive & Social Medicine (by K Park) ✓ Epidemiology (by Leon Gordis) ✓ National Health Programs of India National Policies and Legislations Related to Health (by J. Kishore). ✓ Chapter 53: Public Health Surveillance: A Tool for Targeting and Monitoring Interventions. Available at: https://www.ncbi.nlm.nih.gov/books/NBK11770/pdf/Book shelf_NBK11770.pdf ✓ The CDC Field Epidemiology Manual (by Sonja A. Rasmussen and Richard A. Goodman) ✓ Control of Communicable Diseases Manual (20th Edition)

Course le	Course learning outcomes							
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)					
Unit 1	Introduction to Epidemiology	 Define Epidemiology. (C1) Illustrate the applications of Epidemiology in public health research. (C2) Explain the key terms used in Infectious disease occurrence such as - Outbreak, Epidemic, Cluster, Pandemic, Endemic, Hyperendemic, Sporadic, Epizootic, and Enzootic etc. (C2) Explain Descriptive Epidemiology. (C2) Explain Analytical Epidemiology. (C2) 	2					
Unit 2	Pandemics	 Explain John Snow's work and his contribution to public health. (C2) Examine the history of major pandemics. (C4) Black Death (Plague) Spanish flu COVID-19 	3/3 = 6					



	T		
		 Analyse the factors influencing the spread of pandemics. (C4) 	
Unit 3	Basic concepts in Infectious disease epidemiology	 What is infection and disease? (C1) Define epidemiological triad (Agent, host, and environmental factors). (C1) Explain Iceberg phenomenon in infectious disease. (C2) What are tools of measurements (Rate, ratios, and proportions)? (C1) Summarize measures of disease frequency (Prevalence and Incidence). (C2) Examine and analyse the determinants of health and disease in a population. (C4) 	2
Unit 4	Infectious disease dynamics	 Outline the modes of disease transmission. (C2) Explain portal of entry and exit. (C2) Define incubation period. (C1) Define period of communicability. (C1) Define reproductive number. (R0) (C1) Who are Super spreaders? (C1) How Contact tracing is carried out? (C1) Outline Isolation and quarantine mechanisms. (C2) How to break the chain of transmission in infectious diseases? (C1) Elaborate on hand and respiratory hygiene practices. (C6, A3) 	3
Unit 5	Epidemiological study designs	 Explain epidemiological study designs. (C2) Cross-sectional study Case-control study Cohort study 	5



Compare advantages and disadvantages of various study designs. (CS) Measure strength of association. (CS) Analyse and interpret odds ratio, relative risk and attributable risk. (CS) Define public health surveillance. (C1) Outline different types of surveillance. (C2) Active Passive Sentinel Compare syndromic and disease specific surveillance. (C2) Explain Integrated Disease Surveillance System. (C2) Explain National Vector Borne Disease Control Program. (C2) List the attributes of a good surveillance in evidence-based decision making. (C5) Analyse global surveillance programmes. (FluNET, DenNET, and GLASS). (C4) Outline public health infrastructure in India. (C2) Analyse different health care systems in India (Public, private, PPP model). (C4) Importance of Public health workforce for communicable disease surveillance in India. (C5) Evaluate International Health Regulation (2005) and list of notifiable diseases. (C5) Evaluate International Health Regulation (2005) and list of notifiable diseases. (C5) Explain fundamentals of investigating an outbreak. (C2)		T		T
Analyse global surveillance programmes. (FluNET, DenNET, and GLASS). (C4) Outline public health infrastructure in India. (C2) Analyse different health care systems in India (Public, private, PPP model). (C4) Health care system in India Workforce for communicable disease surveillance in India. (C5) Evaluate International Health Regulation (2005) and list of notifiable diseases. (C5) Unit 8 Outbreak Analyse global surveillance programmes. (FluNET, DenNET, and GLASS). (C4) Importance of Public health workforce for communicable disease surveillance in India. (C5) Evaluate International Health Regulation (2005) and list of notifiable diseases. (C5)	Unit 6		disadvantages of various study designs. (C5) • Measure strength of association. (C5) • Analyse and interpret odds ratio, relative risk and attributable risk. (C5) • Define public health surveillance. (C1) • Outline different types of surveillance. (C2) - Active - Passive - Sentinel • Compare syndromic and disease specific surveillance. (C2) • Explain Integrated Disease Surveillance System. (C2) • Explain National Vector Borne Disease Control Program. (C2) • List the attributes of a good surveillance mechanism. (C4) • Influence of disease surveillance in evidence-based	4
Unit 7 Health care system in India Unit 7 Health care system in India Explain fundamentals of Outline public health infrastructure in India. (C2) Analyse different health care systems in India (Public, private, PPP model). (C4) Importance of Public health workforce for communicable disease surveillance in India. (C5) Evaluate International Health Regulation (2005) and list of notifiable diseases. (C5)		M.	 Analyse global surveillance programmes. (FluNET, DenNET, 	
Unit 8	Unit 7		 Outline public health infrastructure in India. (C2) Analyse different health care systems in India (Public, private, PPP model). (C4) Importance of Public health workforce for communicable disease surveillance in India. (C5) Evaluate International Health Regulation (2005) and list of 	2
	Unit 8		•	4



		 What is an Outbreak? Why should we investigate an outbreak? (C1) Illustrate the steps of outbreak investigation. (C2) Design and develop an outbreak investigation kit. (C6) How to write a Single overriding communication objective (SOCO) statement? (C1) 	
Unit 9	How to conduct a scientific literature search	 Make use of database such as Scopus, Web of Science, PubMed, Medline, Cochrane library, and Google scholar. (C3) Utilize Boolean operators (AND, OR, NOT or AND NOT) for Literature search. (C3) Utilize advanced search features in PubMed (Mesh terms). (C3) Critical Appraisal of a Research Article. (C5) 	2
Unit 10	Epi Info™ 7 Exercise	Analyse the survey data collected from a foodborne outbreak investigation. (Salmonella outbreak tutorial is used to teach the Epi Info™ 7 software to the students and to illustrate how the program can be used to gather, analyse, and present data). (C4, P3)	5 (Tutorials)
Unit 11	Case studies in Applied Epidemiology	Case study- 1 (CDC, No. 401-303) Oswego – An Outbreak of Gastrointestinal Illness Following a Church Supper -Define the terms "cluster," "outbreak," and "epidemic". (C1) -List the steps in the investigation of an outbreak. (C1) -Interpret, draw, and describe the value of an epidemic curve. (C2) -Compare and calculate food-specific attack rates to identify possible vehicles. (C2)	5 (Tutorials)



	-List reasons for investigating an	
	outbreak that has apparently ended.	
	(C4)	
	Case study- 2 (CDC, No. 941-903)	
	Surveillance for E. coli O157:H7	
	Information for Action	
	-Discuss the process and criteria for	
	placing a disease or condition on a	
	state or national notifiable disease list.	
	(C6)	
	-List the categories of information that	
	should be included in a surveillance	5 (Tutorials)
	instrument. (C4)	
	-Interpret and summarize surveillance	
	data. (C5)	
	-Discuss and recognize difficulties in	
	balancing public health concerns with	
	consumer and industry considerations	
	in emerging issues. (C6)	





Manipal Institute of Virology

CURRICULUM

Name of the Program	MSc Clinical Virology				
Course Title	Molecular Virology and Bioinformatics				
Course Code	MIV504				
Academic Year	2022-2024				
Semester	II				
Course credits	4				
Course Prorequisite	First class/CGPA 6.5 at UG level (BSc Life				
Course Prerequisite	Sciences/Health Sciences)				
	The course will offer in-depth knowledge about				
Course Synopsis	nucleic acid extraction, different types of PCR and				
	their principles, basic bioinformatic analysis.				
	CO 1 : Explain the various methods of nucleic acid				
	extraction. (C2, P2)				
	CO 2 : Demonstrate knowledge of application and				
	interpretation of PCR. (C3, P2)				
	CO 3: Interpret the use of bioinformatic tools in				
Course Outcomes	molecular biology. (C2, P2)				
	CO 4: Recall methods of sequencing and				
	molecular cloning. (C1)				
	CO 5: Explain principle, instrumentation and				
	analytical applications of Western blotting. (C2)				
,0 V	CO 6: Explain principle, instrumentation and				
	analytical applications of Microarray. (C2)				

Map	Mapping of COs to POs													
	РО	PO	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО
COs	1	2	3	4	5	6	7	8	9	10	11	12	13	14
CO 1	✓										✓			
CO 2	✓	✓	✓	✓	✓				✓	√	√			
CO 3	✓		✓		✓	✓				✓	✓			
CO 4	√			√		√				✓	✓			
CO 5	√		√	V	√	√		√		✓	√			
CO 6	✓		✓	✓	✓	✓		✓		√	√			



	LEARNING STRATEGY	CONTACT HOURS	SLT	
	Lecture	35	105	
	Seminar	7	21	
	Small Group 7 21			
	Discussion (SGD)			
	Self-directed learning	5	15	
	(SDL)			
Learning Strategies, Contact Hours	Problem Based	6 18		
and Student Learning Time (SLT)	Learning (PBL)			
	Case Based Learning	-	\ <u>-</u>	
	(CBL)			
	Clinic	-	-	
	Practical	-	-	
	Revision	- /-	-	
	Assessment	3	-	
	TOTAL	63	180	
	FORMATIVE	SUMMAT	TVE	
Assessment Methods	Assignment	Mid seme	ster exam	
Assessment Methods	Student presentation	End seme	ster exam	
	Group discussion			

Mapping of assessment with COs						
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5	CO 6
Assignments			✓	✓		
Student presentations			✓		✓	✓
Group discussion	✓	✓	✓			
Mid-semester examination	✓	✓				
End-semester examination	✓	✓	✓	✓	✓	✓
Practical examination	×	×	×	×	×	×

Student feedback on Course and Course master.
 Molecular Cloning- A laboratory manual: Sambrook and Russell
 Molecular Cloning- A laboratory manual: Green and Sambrook
 Gene cloning and DNA analysis- An introduction: TA Brown
 Essential bioinformatics – Jin Xiong Bioinformatics sequence and genome analysis: David W Mount



Course le	arning outcomes		
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)
Unit 1	Nucleic acid amplification and sequencing	 Explain the principle and various methods of nucleic acid extraction. (C2) Demonstrate in-house and kit-based method of nucleic acid extraction. (C2, P2) Outline different types of PCR and explain the principles. (C2) Demonstrate conventional PCR, electrophoresis, realtime PCR and quantitative PCR. (C2, P2) Explain the methods of nucleic acid sequencing. (C2) 	12/6 = 18
Unit 2	Molecular diagnostic Virology	 Apply PCR tests for diagnosis of viral infections. (C3, P3) Interpret results of PCR. (C2, P2) 	5/2 = 7
Unit 3	Phage library and Reverse Genetics	 Explain phage library and its application. (C2) Explain reverse genetics and its application. (C2) 	2/1 = 3
Unit 4	Prokaryotic and Eukaryotic gene expression	Explain the methods of prokaryotic and eukaryotic gene expression. (C2)	2/1 = 3
Unit 5	Cloning and cloning Vectors, expression vectors	 Explain cloning and expression vectors. (C2) 	2/1 = 3
Unit 6	Introduction to biological databases	Explain different types biological databases and applications. (C2)	2/1 = 3
Unit 7	Bioinformatics software	Apply bioinformatic software for sequence search, primer and probe	4/2 = 6



		decigning and coguence	
		designing, and sequence alignment. (C3, P2)	
Unit 8	Sequence analysis	• Experiment with nucleic acid sequence data. (C3)	3/1 = 4
Unit 9	Phylogenetics	 Select and utilize different software for phylogenetic analysis. (C3, P2) 	3/1 = 4
Unit 10	Structure based approach in drug design, vaccine targets / virus attachment	 Select and utilize different software for drug design, vaccine targets / virus attachment. (C3, P2) 	2/1 = 3
Unit 11	Western blotting	 Explain principle of Western blotting. (C2) Outline instrumentation. (C2) List analytical applications. (C2) 	2/1 = 3
Unit 12	Microarray	 Explain principle of Microarray. (C2) Outline instrumentation. (C2) List analytical applications. (C2) 	2/1 = 3





Name of the Program	MSc Clinical Virology			
Course Title	Virological Techniques			
Course Code	MIV506			
Semester	II MII,			
Course credits	4			
Academic Year	2022-2024			
Course Prerequisite	First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences)			
Course Synopsis	 This module helps to understand the basic virological techniques. To provide fundamental knowledge of various classical and modern virological techniques as well as the technique used for the propagation and quantification of viruses in a laboratory setting. To provide an understanding of the principles and practical applications of techniques used in biomedical research and clinical diagnostics. 			
Course Outcomes	CO 1: Explain the various methods of isolation			
	and quantification of viruses. (C2, P3)			
	CO 2: Describe the various methods to detect virus growth. (C2) CO 3: Distinguish the advantages and disadvantages of the different quantification and propagation methods. (C4)			
	CO 4: Explain the various assays for			
	neutralization of viruses. (C2, P3)			
	CO 5: Evaluate the techniques for isolation,			
	quantification and neutralization of viruses in lab. (C5, P3)			
	CO 6: Describe various ELISA techniques. (C2, P2) CO 7: Illustrate principle, instrumentation and analytical applications of Chromatography. (C2) CO 8: Demonstrate principle, instrumentation and analytical applications of Spectrophotometry. (C2, P2)			



CO 9: Illustrate principle, instrumentation and analytical applications of Ultracentrifugation. (C2)**Mapping of COs to POs** PO COs 2 3 4 5 8 10 11 12 13 14 6 CO ✓ ✓ ✓ СО ✓ ✓ 2 СО ✓ ✓ CO CO ✓ CO ✓ CO ✓ 7 CO 8 CO **Learning Strategies, Contact Hours LEARNING CONTACT** SLT and Student Learning Time (SLT) **STRATEGY** HOUR 120 Lecture 40 Seminar 15 45 Small 1 3 Group Discussion (SGD) Self-directed 3 9 learning (SDL) Problem Based 1 3 Learning (PBL) Case Based Learning (CBL) Clinic -Practical Revision Assessment 3 TOTAL 63 180 **Assessment Methods FORMATIVE SUMMATIVE** Mid-semester exam Assignment



Student presentation
Group discussion

End semester exam

Mapping of assessment with COs									
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5	CO 6	CO 7	CO 8	CO 9
Assignments		✓					✓	✓	
Student presentations						✓			✓
Group discussion									
Mid-semester examination	✓	✓	✓						
End-semester examination	✓	✓	✓	✓	✓	✓	✓	✓	✓
Practical examination	×	×	×	×	×	×	×	×	×

Feedback Methods	Student feedback on Course and Course master.						
Main Reference	• Diagnostic procedures for Viral, Rickettsial, and Chlamydial						
	Infections- E H Lennette et al.						
	Color Atlas of Virology- Versteeg J.						
	Primer Practical Manual in Classical Viral isolation						
	Techniques- Roshan. J. et al.						
	• Clinical Virology Manual, fourth edition - Specter S et al.						
	Clinical Microbiology procedures- Handbook- Isenberg						
	WHO/CDC Manual						
	 https://www.classcentral.com/course/swayam-analytical- 						
	techniques-13896						
	• Instrumental methods of analysis H.H.Wilard, L.L.Merritt, J						
	A Dean.						

Course learning outcomes								
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)					
Unit 1	Propagation of Viruses - Animal inoculation, Chick - embryo inoculation, cell culture inoculation Detection of Virus growth - Cytopathic effect,	methods for isolation of viruses. (C3, P1)	6/2 = 8					
Unit 2	Quantitation of Viruses	 Describe the various methods for quantification of viruses. (C3, P1) 	5/3 = 8					



	T -		1
	-TCID ₅₀ , Hemagglutination assay, -Plaque assay	 Discuss the advantages and disadvantages of the various quantification methods. (C2) Practice the different virus quantification methods. (C3, P3) Describe the principle and procedure for Hemagglutination assay. (C3, P3) Explain the principle and procedure for Plaque assay. (C3, P3) Discuss the principle and procedure for TCID₅₀ assay. (C3, P3) 	
	Virus Neutralization Assay- Microneutralization Assay, Plaque reduction neutralization Assay	 Describe the principle and procedure for Microneutralization Assay. (C2, P3) Illustrate the applications of virus neutralization assay. (C3) Describe the principle and procedure for Plaque reduction neutralization Assay. (C3, P3) 	5/3 =8
Unit 3	Hemagglutination Inhibition Assay (HAI)	 Describe the principle and procedure for Hemagglutination. inhibition assay. (C2, P3) Illustrate the applications of HAI. (C3) 	4/4 = 8
Unit 4	Complement fixation Assay	 Describe the technique of compliment fixation. (C2) 	3/1 = 4
Unit 5	Virus Interference Assay	 Describe the technique of Virus Interference Assay. (C3) 	· ·
Unit 6	Immunofluorescence assay (IFA)	 Illustrate the steps for developing Immunofluorescence assay for the detection of IgG antibodies to measles virus. (C3, P3) 	4/3 = 7



	Γ	Т	
Unit 7	Enzyme-Linked Immunosorbent Assay (ELISA)	 Describe the principle of ELISA technique. (C2) Explain types of competitive and non-competitive ELISAs using appropriate diagrams. (C2, P3) Illustrate the applications of ELISA. (C3, P3) 	4/3 = 7
Unit 8	Spectrophotometry	 Demonstrate the principle of Spectrophotometry. (C3) Outline instrumentation. (C3) List analytical applications. (C3) 	1/1 = 2
Unit 9	Ultra-centrifugation	 Illustrate the principle of Ultracentrifugation. (C3) Outline instrumentation. (C3) List analytical applications. (C3) 	1/1 = 2
Unit 10	Chromatography	 Illustrate the principle of Chromatography. (C3) Outline instrumentation. (C3) List analytical applications. (C3) 	1/1 = 2





Name of the Program	MSc Clinical Virology				
Course Title	Analytical Tools (Application of GIS and				
	Biostatistics)				
Course Code	MIV508				
Academic Year	2022-2024				
Semester	II O				
Course credits	2				
Course Prerequisite	First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences)				
	 To understand basic concepts of Geographical Information System (GIS). To know the different types of data formats used in GIS. 				
	3. To understand the methods of spatial analysis and its interpretations.				
Course Synopsis	 4. To support the students to utilize and apply GIS concepts in viral disease epidemiology. 5. This module introduces to the concepts in Biostatistics. 				
	6. To understand the applications of biostatistics in research and public health.7. To introduce statistical packages for data analysis in MS Excel, SPSS, and Epi Info.				
MRII.	CO 1: Outline the basic concepts of GIS, History of mapping, geo co-ordinate system and software. (C2) CO 2: Classify different types of data formats used in GIS. (C2)				
Course Outcomes	CO 3: Analyse the distribution, dependency, and interaction between spatial data. (C4) CO 4: Interpret the output of spatial analysis performed in GIS. (C2) CO 5: Illustrate the applications of GIS in viral disease epidemiology. (C2)				



CO 6: Outline basic terminologies used in biostatistics. (C2)

CO 7: Explain basic concepts in biostatistics— Variables, types of variables, scales of measurements, descriptive statistics, and inferential biostatistics, etc. (C2)

CO 8: Discover various methods of data collection. (C4)

CO 9: Summarize various methods of sample size calculation. (C2)

CO 10: Identify the variables in a dataset and classify variables. (C3)

CO 11: Create graphs & diagrams using datasets. (C6)

CO 12: Use statistical software for data analysis and interpretation. (C4, P3)

Mapp	Mapping of COs to POs													
	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО
COs	1	2	3	4	5	6	7	8	9	10	11	12	13	14
CO 1	✓									✓	✓		✓	
CO 2	✓		✓	✓						√	✓			
CO 3	✓		✓			✓			✓	✓	✓			
CO 4	✓			✓					✓	✓	✓			
CO 5	✓		✓			✓			✓	✓	✓			
CO 6	✓										✓	✓		
CO 7	✓			√						✓	✓	✓		
CO 8	✓		. \$	/	✓	þ		✓	✓	✓	✓	✓		
CO 9	✓		~	~	✓				✓	✓	✓	✓		
CO 10	✓			√	✓					√	√	✓		
CO 11	✓		✓	✓	✓				✓	✓	✓	✓		
CO 12	✓		✓	✓	✓				✓	✓	✓	✓		

Learning Strategies, Contact Hours and Student Learning Time (SLT)

LEARNING	CONTACT	SLT
STRATEGY	HOURS	
Lecture	10	30
Seminar	3	9
Small Group	2	6
Discussion (SGD)		
Self-directed	6	18
learning (SDL)		



	Problem Based Learning (PBL)	1	3
	Case Based Learning	2	6
	(CBL)		
	Clinic	-	-
	Practical	6	18
	Revision	-	-
	Assessment	1	-
	TOTAL	31	90
	FORMATIVE	SUMMA	TIVE
Assessment Methods	Assignment	Mid sem	nester exam
Assessificiti Metilous	Student presentation		
	Group discussion		

Mapping of assessment with COs												
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5	C O 6	C O 7	C O 8	C O 9	C O 10	C O 11	C O 12
Assignments			✓		V						✓	
Student presentations		✓	✓ .	V	1		✓	✓	✓	✓		
Group discussion	✓					✓						✓
Mid-semester examination	✓	✓	~)		✓	✓	✓	✓			
End-semester examination	×	×	×	×	×	×	×	×	×	×	×	×
Practical examination	×	×	×	×	×	×	×	×	×	×	×	×

Feedback Methods	Student feedback on Course and Course master.					
Reference Materials	 GIS tutorial for Health - Kristen S. Kurland Understanding GIS - Christian Harder Fundamentals of GIS - Nick Santos (UC DAVIS) An Introduction to Biostatistics. A manual for students in health sciences. (P.S.S. Sundar Rao & J. Richard) Principles and Practice of Biostatistics. (B. Antonisamy) IBM SPSS Statistics v20, User Manual. 					
	Getting started with EpiInfo 7, User guide from CDC					



Course le	arning outcomes		
Content	Topics	Learning Outcomes	Hours (Lectures/ Tutorials)
Unit 1	Basic concepts of GIS	 Define Geographical information system. (C1) Recall the history of mapping and John snow's work on Cholera. (C1) Explain the fundamental of Geo-Positioning System (GPS). (C2) List of different GIS software available. (C1) Outline the applications of GIS in public health. (C2) 	1
Unit 2	GIS Data formats and Vector data	 Explain Vector, Raster, shapefile, excels, textfile, etc., with examples. (C2) Demonstrate the user interface of ArcGIS software package. (C2) Explain point data, line data and polygon data with examples. (C2) 	1/1 = 2
Unit 3	Raster Data	 Explain raster data with example. (C2) Demonstrate the base map, surface map and thematic map. (C2) 	1/1 = 2
Unit 4	Common tools used in ArcGIS and Attribute table	 List common tools used in ArcGIS. (C1) Demonstrate the "Add data" tool, "Folder connection" and "Layers". (C2) Demonstrate the "Edit", "Selection", "Join", "draw", "Display XY", "Export" and "Merge" tools. (C2) Demonstrate the "Properties" tab. (C2) Define Attribute table. (C1) Outline the ways to create, add and edit columns in attribute table. (C2) 	1+1 = 2



	T		
Unit 5	Symbology and label	 Define "Symbology" tab. (C1) Demonstrate "Quantitative" symbology (Graduated colours, symbols, and Dot density). (C2) Demonstrate "Chart" symbology (Pie, Bar, Stack). (C2) Demonstrate "Multiple" attribute symbology. (C2) Demonstrate "Label" feature. (C2) 	1/1 = 2
Unit 6	Spatial analysis	 Demonstrate health data visualization such as choropleth map, dot-density map, etc. (C2) Explain "Spatial analyst" toolbox. (C2) Explain "Spatial statistics" toolbox. (C2) Analyse hotspot. (C4) Analyse cluster. (C4) 	2/1 = 3
Unit 7	Application of GIS in Public health	 Explain the application of GIS in epidemiology and disease surveillance. (C2) Explain the application of GIS Prediction model, risk and response assessment. (C2) Analyse important viral diseases of India using GIS. (C4) 	2/1 = 3
Unit 8	Introduction to Biostatistics	 Define Biostatistics and key terms used in biostatistics. (C1) Applications of Biostatistics in research. (C3) 	1
	Data collection methods	 What is Data, dataset, and data table? (C1) Explain Variables and types of variables. (C2) Understand various methods of data collection (Qualitative and Quantitative). (C2) Develop research questionnaire. (C6) 	1



			T
		 Outline Scales of measurements. (C2) Understanding frequency distribution and class intervals. (C2) 	
Unit 9	Measures of central tendency & measures of dispersion	 Explain Measures of central tendency (Mean, median, mode). (C2) Explain measures of dispersion (standard deviation, variance, quartiles, range, Inter quartile range). (C2) Solve mathematical problems using datasets. (C3) 	
Unit 10	Sampling/ Sampling methods	 Explain principal concepts in sampling. (C2) What is population and sample, parameter, and statistics? (C1) Compare probability and non-probability sampling techniques? (C2) What is multistage sampling technique? (C1) Explain the advantages of various sampling techniques? (C2) 	1
Unit 11	Normal distribution	 Define normal distributions. (C1) Properties of normal distributions. (C2) What is Empirical rule? (C1) Explain Reference limits. (C2) What is Standard normal distribution? (C1) What is Z-score? (C1) Solve mathematical problem using datasets. (C3) 	1
Unit 12	Representation of data	 Explain various methods of representation of data (text, tabulation and graphical). (C2) Construction of various type of graphs and interpretation. (C6) 	1



	1		T
		Identify the suitable type of graphical representation for a	
		 given set of data. (C3) Elaborate on the different approaches used in sample size 	
Unit 13	Sample size determination	calculation. (C6)Analyse dataset and run statistical tests. (C4)	1
	Correlation and regression	 Outline correlation and regression methods. (C2) Classify and compare positive, negative, and neutral correlation. (C2) Calculate and interpret correlation coefficient. (C5) Analyse dataset and run binary logistic regression analysis. (C4) 	1
Unit 14	Data management	 Make use of MS Excel, SPSS & Epi info for data management. (C3) Explain data collection, organization, cleaning, storage and archiving, sharing and security, and quality control. (C2) 	1
Unit 15	Hands on training in MS Office	 Summarizing data using MS Excel (C2) fundamentals of MS Excel and spreadsheet Create and modify charts Pivot table and pivot charts Data validation in excel Making Dashboard Tabulation Demonstrate formatting in MS word (C2) Document formatting, paragraphs, headings, page break, page numbering Using templates & styles Table of contents, list of figures, templates 	1/2 = 3



	T		
		Make use of MS PowerPoint for	
		(C3)	
		- Basic tasks	
		 Templates and graphics 	
		 Best practices in making a 	
		scientific presentation	
		Make use of Mendeley reference	
		management software (C3)	
		 Desktop version 	
		- Web importer	
		- Word plugin	
		Outline the fundamentals of using	
		SPSS. (C2)	1 /// .
		Understand the basic workings of	
		SPSS and data entry. (C2)	0, ,
		How to compute variables, recode	
		into new variable? (C1)	
		Organise data using sort cases and	
		select cases. (C3)	
		• Create simple tables and charts	
11.31.46	Hands on	using Chart builder. (C6)	4/2 2
Unit 16	training in SPSS	How to export and import SPSS	1/2= 3
		data? (C1)	
		Perform basic statistical analyses	
		using Descriptive statistics. (C2)	
	(C	Analyse and perform advanced	
	18	statistical tests with SPSS	
		(statistical tests, correlation, and	
		regression analysis). (C4)	
		Demonstrate R statistical software	
		(RStudio and R commander). (C2)	





Name of the Program	MSc Clinical Virology
Course Title	Emerging Viral Diseases and Public health
Course Title	response
Course Code	MIVEL510.1
Academic Year	2022-2024
Semester	II
Course credits	3
Course Proroguicite	First class/CGPA 6.5 at UG level (BSc Life
Course Prerequisite	Sciences/Health Sciences)
Course Synopsis	Emerging infectious diseases (EIDs) are serious public health threats globally. An emerging infectious disease is one that either has appeared and affected a population for the first time, or has existed previously but is rapidly spreading, either in terms of the number of people getting infected, or to new geographical areas. Many EIDs are zoonotic in origin, which means that the disease has emerged from an animal and crossed the species barrier to infect humans. Humans may often have little or no natural immunity to EIDs, so their impact on health, society and the economy, are difficult to predict. • This course will help in the understanding of emerging and re-emerging viral diseases of national and international concern. • This course will provide a description of the type, severity, transmission and spread of infectious diseases with an explanation of risk
	infectious diseases, with an explanation of risk factors and symptoms of infection in humans. CO 1: Explain emerging and re-emerging viral
	diseases. (C2)
	CO 2: Explain reasons for emergence and re-
Course Outcomes	emergence of infectious viral diseases. (C2)
Course Outcomes	CO 3: Outline risk distribution of global emerging
	infectious diseases. (C2)
	CO 4: Explain WHO's response to International
	Health Emergencies. (C2)



CO 5: Illustrate International Health Regulations. (C2) **Mapping of COs to POs** РО COs 14 2 8 9 10 11 12 13 co CO ✓ ✓ ✓ co **√ √** ✓ CO 4 ✓ **√** CO 5 **LEARNING STRATEGY** CONTACT SLT **HOURS** Lecture 30 90 15 Seminar 5 Small Group 3 9 Discussion (SGD) Self-directed learning 7 21 (SDL) **Learning Strategies, Contact Hours** Problem Based and Student Learning Time (SLT) Learning (PBL) Case Based Learning (CBL) Clinic Practical Revision Assessment 1 46 **TOTAL** 135 **FORMATIVE SUMMATIVE** Assignment Mid semester exam **Assessment Methods** Student presentation Group discussion

Mapping of assessment with COs					
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5
Assignments	✓		✓		
Student presentations				✓	
Group discussion			✓	✓	✓
Mid-semester examination	✓	✓	✓		
End-semester examination	×	×	×	×	×
Practical examination	×	×	×	×	×



Feedback Methods	Student feedback on Course and Course master.					
Reference Materials	 https://www.ijmr.org.in/article.asp?issn=0971-5916;year=2019;volume=149;issue=4;spage=447;epage=467;aulast=Mourya https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3767269/ https://www.who.int/zoonoses/diseases/en/ https://www.pnas.org/content/97/23/12411 					

Course le	arning outcomes		V/V/
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)
Unit 1	Emerging Viral Diseases	 Summarize emerging infectious diseases in the recent past. (C2) Outline emerging and re-emerging viral diseases in India. (C2) Explain the factors contributing to the emergence of viral diseases. (C2) 	6/8 = 14
Unit 2	Pandemic and epidemic- prone diseases	Understanding the pandemic and epidemic-prone diseases. (C2)	6/4 = 10
Unit 3	International Health Regulations	Illustrate International Health Regulations. (C2)	4/4 = 8
Unit 4	Public health response in Viral disease emergencies	 Summarize WHO's and ICMR's response to national and international health emergencies. (C2) 	5/8 = 13





Name of the Program	MSc Clinical Virology
Course Title	One Health in Virology
Course Code	MIVEL510.2
Academic Year	2022-2024
Semester	II
Course credits	3
Course Prerequisite	First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences)
Course Synopsis	 This module introduces the principles of diseases in the context of socio-ecological systems and global health. It is aimed to provide knowledge and skills in relation to One Health methodologies, transdisciplinary interactions and in using a systems approach.
Course Outcomes	co 1: Understand the One Health concept and approach problem solving using a transdisciplinary methodology in Virology. (C1) CO 2: Understand the origin, context and drivers of viral diseases at the human, animal and environment interface. (C1) CO 3: Evaluate impacts of multi-host infections on human, animal and ecosystem health and economics directly or indirectly, via food, disease vectors or the environment. (C5) CO 4: Develop a One Health systems approach to complex disease issues in monitoring. (C6) CO 5: Critically review published literature. (C5) CO 6: Design a research project. (P5)



	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	PO1	PO1	
Os	1	2	3	4	5	6	7	8	9	10	11	12	3	4	
1 C	√										✓				
) 2	✓		✓					✓	✓		✓				
3	√										√	✓			
0 4	\		√					✓	✓		✓				
0 5	✓		✓	✓				✓	✓		✓				
06	✓		✓	✓		✓			✓		✓		✓		
							LE/	ARNIN	G STF	RATEGY	CC	NTACT		SLT	
											H	IOURS			
							Lec	ture				15		45	
							Ser	ninar				12		36	
				Small Group					6		18				
							Dis	Discussion (SGD)							
							Sel	Self-directed learning				6		18	
							(SD	L)							
arni	ng Sti	rategi	es, Co	ontact	Hour	`S	Pro	Problem Based				6		18	
d St	uden	t Lear	ning	Time	(SLT)		Lea	rning	(PBL)						
							Cas	se Bas	ed Le	arning		-		-	
							(CE	SL)							
							Clir	nic				-		-	
			Practical							-		-			
						Rev	Revision				-		-		
							Assessment					1		-	
							TO	TAL				46		L35	
							FO	RMAT	IVE			SUMMA	TIVE		
202	ment	t Metl	hods				Ass	signme	ent			Mid sen	nester	exam	
·C33	,,,, C ,,,	LIVIELI	ious							ntation					
							Gra	oup di	courci	on					

Mapping of assessment with COs						
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5	CO 6
Assignments						✓
Student presentations		✓	✓	✓	✓	
Group discussion	✓	✓	✓	✓	✓	
Mid-semester examination	✓	✓	✓	✓		
End-semester examination	*	×	×	×	×	×
Practical examination	*	*	*	×	×	×



Feedback Methods	Student feedback on Course and Course master.						
Reference Materials	 ✓ One Health: People, Animals and Environment by Atlas and Maloy ✓ Sustaining Global Surveillance and Response to Emerging Zoonotic Diseases – Institute of Medicine and National Research Council ✓ Basic epidemiology: WHO http://whqlibdoc.who.int/publications/2006/9241547073 e ng.pdf ✓ Lueddeke G. Survival: One Health, One Planet, One Future. Routledge. September 2018 ✓ The CDC Field Epidemiology Manual: https://www.cdc.gov/eis/field-epimanual/chapters/Describing-Epi-Data.html 						

Course learning outcomes							
Content	Topics Learning Outcomes		Hours (Lectures/Tutorials)				
Unit 1	Foundations of One Health	 An introduction to One Health. (C2) The integration of One Health in Virology. (C2) 	4/4 = 8				
Unit 2	Emergence of Infectious Diseases	 Introduction to disease agents for One Health. (C2) Understanding the cause of emergence of viral infections. (C2) Outbreaks and their control. (C2) 	4/4 = 8				
Unit 3	Introduction to One Health Epidemiology and Surveillance	 Understand the basic principles of epidemiology in One Health. (C2) Understand environment and ecosystem health. (C2) Understand surveillance involved in One Health. (C2) 	4/4 = 8				
Unit 4	Public Health and One Health	 Understand public health policy and socioeconomic principles of One Health. (C2) Understand the role of national government and international organisations in implementing public health policy. (C2) 	3/3 = 6				





Name of the Program	MSc Clinical Virology						
Course Title	Practical II (Molecular and Virological						
Course Title	Techniques)						
Course Code	MIV512						
Academic Year	2022-2024						
Semester	II O						
Course credits	3						
Course Prerequisite	First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences)						
Course Synopsis	 The course will offer in-depth knowledge about nucleic acid extraction, different types of PCR and their principles, basic bioinformatic analysis. This module also helps to understand the basic virological techniques. To provide fundamental knowledge of various classical and modern virological techniques used for propagation and quantification of viruses in laboratory setting. 						
Course Outcomes	Molecular techniques: CO 1: Explain the various methods of nucleic acid extraction. (P2) CO 2: Demonstrate knowledge of application and interpretation of PCR. (P2) CO 3: Interpret the use of bioinformatic tools in molecular biology. (P2) Virological techniques: CO 4: Explain the various methods of isolation and quantification of viruses. (P3) CO 5: Explain the various assays for neutralization of viruses. (P3) CO 6: Evaluate the techniques for isolation, quantification and neutralization of viruses in lab. (P3)						



								CO 7:	Descr	ibe va	rious I	ELISA tec	hnique	s. (P2)	
Мар	ping	of CO	s to P	Os (N	lolecu	lar te	chni	ques)							
	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	
COs	1	2	3	4	5	6	7	8	9	10	11	12	13	14	
СО	\checkmark										✓				
CO CO	√	√	√	√	√				✓	√	✓				
2	V	•	•	•	•				•	"	•				
СО	√		√		√	√				√	√				
3															
Map	ping	of CO	s to P	Os (Vi	rolog	ical te	echn	iques)							
	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	
COs	1	2	3	4	5	6	7	8	9	10	11	12	13	14	
СО	✓					✓		✓			√		/		
4 CO	▼					V		-			-		•	-	
5	•					✓					✓				
СО	✓											S			
6						✓			✓		✓				
СО	\checkmark			√	✓	✓									
7				V	v	•					✓				
										NING		CONTACT		SLT	
									STRA	TEGY		HOUR	S		
								Lect	ure			-		-	
								Seminar			-		-		
								Small Group				-		-	
								Discussion (SGD)					_		
								Self-directed learning (SDL) Problem Based				_			
.earni	ing St	rateg	ies. C	ontac	t Hou	rs and	d							 	
Stude	_	_				/						-		_	
			,	. (02,				Learning (PBL)							
								Case Based Learning				-		-	
								(CBL)							
								Clini	С			-		-	
								Prac	tical			90		270	
								Revi	sion			-		-	
								Assessment TOTAL				2		-	
												92		270	
- 1/1/2·					MATI	VE		SUMM	IATIVE						
								-	ssme			End se		exam	
Accece	smen	t Mot	hode							111 01	}	Liid 3C		CAUIII	
Assessment Methods			Record books												
							I	I hool	(S						

Feedback Methods	Student feedback on Course and Course master.
Reference Materials	Refer to MIV504 and MIV506



Mapping of assessment with COs									
Nature of assessment	CO 1	CO 2	соз	CO 4	CO 5	CO 6	CO 7		
Record books, technical skills, conduct in lab	✓	✓	✓	✓	✓	✓	✓		
Internal assessment	✓	✓	✓	✓	✓	✓	√		
Assignments	×	×	×	×	×	×	×		
Student presentations	×	×	×	×	×	×	×		
Group discussion	×	×	×	×	×	×	×		
Mid-semester examination	×	×	*	×	×	×	×		
End-semester examination	✓	✓	√	✓	✓	✓_	✓		

Course learni			
Content	Topics	Learning Outcomes	Practicals
Unit 1 (MIV504)	Nucleic acid amplification and sequencing	 Explain the principle and various methods of nucleic acid extraction. (C2) Demonstrate in-house and kit-based method of nucleic acid extraction. (C2, P2) Outline different types of PCR and explain the principles. (C2) Demonstrate conventional and real-time PCR. (C2, P2) Explain the methods of nucleic acid sequencing. (C2) 	8
Unit 2 (MIV504)	Molecular diagnostic Virology	 Apply PCR tests for diagnosis of viral infections. (C3, P3) Interpret results of PCR. (C2, P2) 	7
Unit 4 (MIV504)	Prokaryotic and Eukaryotic gene expression	Explain the methods of prokaryotic and eukaryotic gene expression. (C2)	2
Unit 5 (MIV504)	Cloning and cloning Vectors, expression vectors	Explain cloning and expression vectors. (C2)	2



		Explain different types	
Unit 6 (MIV504)	Introduction to biological databases	biological databases and applications. (C2)	2
Unit 7 (MIV504)	Bioinformatics software	Apply bioinformatic software for sequence search, primer and probe designing, and sequence alignment. (C3, P2)	3
Unit 8 (MIV504)	Sequence analysis	• Experiment with nucleic acid sequence data. (C3)	2
Unit 9 (MIV504)	Phylogenetics	 Select and utilize different software for phylogenetic analysis. (C3, P2) 	2
Unit 10 (MIV504)	Structure based approach in drug design, vaccine targets / virus attachment	 Select and utilize different software for drug design, vaccine targets / virus attachment. (C3, P2) 	2
Course learning	ng outcomes (Virologica	l techniques)	
Content	Topics	Learning Outcomes	Hours (Practicals)
Unit 1 (MIV506)	Propagation of Viruses - Animal inoculation, Chick - embryo inoculation, cell culture inoculation Detection of Virus growth - Cytopathic effect,	 Discuss the various methods for isolation of viruses. (C3, P1) Explain the advantages and disadvantages of various virus isolation methods. (C2) Demonstrate virus isolation using cell culture. (C3, P3) Identify the various cytopathic effects of viruses. (C3, P1) 	10
Unit 2 (MIV506)	Quantitation of Viruses -TCID ₅₀ , Hemagglutination assay, -Plaque assay	 Describe the various methods for quantification of viruses. (C3, P1) Discuss the advantages and disadvantages of the various quantification methods. (C2) Practice the different virus quantification methods. (C3, P3) Describe the principle and procedure for 	10



	Virus Neutralization	 Hemagglutination assay. (C3, P3) Explain the principle and procedure for Plaque assay. (C3, P3) Discuss the principle and procedure for TCID₅₀ assay. (C3, P3) 	10
	Assay- Microneutralization Assay, Plaque reduction neutralization Assay	 Describe the principle and procedure for Microneutralization Assay. (C2, P3) Illustrate the applications of virus neutralization assay. (C3) Describe the principle and procedure for Plaque reduction neutralization Assay. (C3, P3) 	
Unit 3 (MIV506)	Hemagglutination Inhibition Assay (HAI)	 Describe the principle and procedure for Hemagglutination inhibition assay. (C2, P3) Illustrate the applications of HAI. (C3) 	10
Unit 6 (MIV506)	Immunofluorescence assay (IFA)	• Illustrate the steps for developing Immunofluorescence assay for the detection of IgG antibodies to measles virus. (C3, P3)	10
Unit 7 (MIV506)	Enzyme Linked Immunosorbent Assay (ELISA)	 Describe the principle of ELISA technique. (C2) Explain types of competitive and non-competitive ELISAs using appropriate diagrams. (C2, P3) Illustrate the applications of ELISA (C3, P3) 	10





Manipal Institute of Virology

CURRICULUM

Learning Strategies, Contact Hours and			Hours		LEAR		CONTACT SLT							
CO 2	✓		✓	5		✓					✓		✓	
CO 1	✓					✓			✓		✓			
COs	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
Мар	ping o	f COs	to POs	1						T		T	1	
							pra	practices. (C2)						
							CO	CO2: Recall and relate good laboratory						
							virology laboratory. (C2)							
Cours	e outo	omes					CO1: Outline the workflow in a diagnostic							
Cours	e Synd	opsis					I	This course will provide an overall view of the work flow in a diagnostic virology laboratory.						
C	- C							ences/l	-					- £ + l-
Cours	e Prer	equisi	te					t class				i leve	el (BSo	: Life
	f credi						2							
Seme							Ш				1			
	emic Y	ear						2-202	4					
Cours	e Cod	е					M۱۱	/514)
Cours	e Title	1					Lab	orato	y Rota	tion	- I			
Name of Program						MS	MSc Clinical Virology							

Learning Strategies, Contact Hours and Student Learning Time (SLT)	LEARNING STRATEGY	CONTACT HOUR	SLT
Student Learning Time (SLT)	JINAILUI	HOOK	
	Lecture	-	-
	Seminar	-	-
	Small Group	-	-
	Discussion		
M_{I} ,	(SGD)		
	Self-directed	-	-
	learning (SDL)		
	Problem Based	-	-
	Learning (PBL)		
	Case Based	-	-
	Learning (CBL)		
	Laboratory	60	180
	Practical	-	-



	Revision		-	-					
	Assessment 1		Assessment 1		Assessment 1		Assessment 1		
	TOTAL	(61	180					
Assessment Methods	FORMATIVE			SUMMATIVE					
	Assessment of Lab posting Log Books		-	-					
	Internal								
	Assessment- MCQs	;							

Foodbook Mothodo	Cturd out foodbook on Course and Course months	
Feedback Methods	Student feedback on Course and Course master.	

Posting	Sections	Learning Outcomes	Hours
1	Serology	 Observe and understand the workflow in the diagnostic laboratory. 	60 hours (12 hours in each section)
2	Sample processing and Extraction	 Observe and understand the initial processing of clinical samples. 	355000.1,
3	Molecular Diagnostics	 Observe and understand the principle, methodology and 	
4	Tissue culture	technique of the different tests	
5	Sample reception, Barcoding, Sample Storage and Decontamination	 (serological, molecular and cell culture based) performed routinely in a diagnostic laboratory. Observe sample reception, barcoding and sample storage processes. Observe and understand the protocols of decontamination 	
		and discarding of biomedical waste.	



THIRD SEMESTER





Manipal Institute of Virology

РО

COs

РО

2

РО

3

РО

4

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6

РО

7

Name of the Program	MSc Clinical Virology
Course Title	Clinical & Diagnostic Virology-I
Course Code	MIV601
Academic Year	2022-2024
Semester	III
Course credits	4
Course Prerequisite	First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences)
Course Synopsis	 This module introduces to the principles of clinical and diagnostic virology. Differential diagnosis of various clinical presentations. To understand the syndromic approach towards diagnosis of viral infections. Understanding the diagnostic algorithms of various viral infections. Performing various tests required to diagnose various viral infections.
Course Outcomes	co 1: Understand the basic concepts of clinical and diagnostic virology. (C1) co 2: Interpret & infer information from patient case sheets from hospitals. (C2) co 3: Outline the clinical features of various clinical infections. (C2) co 4: Identify the differential diagnosis of various clinical presentations. (C3) co 5: Construct diagnostic algorithms of various viral infections. (C6) co 6: Demonstrate laboratory skills to perform various diagnostic tests. (C2, P5) co 7: Evaluate and discuss case studies in clinical virology. (C5)



РО

8

РО

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РО

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РО

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РО

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РО

13

PO

14

СО	✓										✓			
CO	√		✓					/	✓		✓			
2	V		•					•	•		•			
CO 3	✓										✓			
CO 4	√		√					✓	✓		√			
CO 5	√		√	√				✓	√		√			
CO 6	√		√	√		√			√		✓		√	
CO 7	√		√	√				✓	√		√			
						ARNI TRATE		(CONTACT		SLT			
							L	ecture				30		90
							S	emina	r			12		36
							Small Group Discussion (SGD)					6		18
							S			learnin	g	4		12
	_	_		ontac Time	t Hou (SLT)	rs	P	Problem Based Learning (PBL) Case Based Learning (CBL)				4		12
							С							
							C	linic				-		-
							Р	ractica	al			-		
							R	evisio	n	-		-		-
								ssessr	nent			3		-
				<u>,c</u>				OTAL				63		180
					J			ORMA				SUMM		
Asses	smen	t Met	hods					ssignr				Mid ser		
		1								entatio	n	End ser	nester	exam
							l I G	iroup (aiscus	sion				

Mapping of assessment with COs											
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5	CO 6	CO 7				
Assignments							✓				
Student presentations		✓	✓	✓	✓		✓				
Group discussion	✓	✓	✓	✓			✓				
Mid-semester examination	✓	✓	✓	✓							
End-semester examination	✓	✓	✓	✓	✓	✓	✓				
Practical examination	×	×	×	×	×	×	×				



Feedback Methods	Student feedback on Course and Course master.							
Reference Materials	 ✓ Lennette's Laboratory diagnosis of viral infections (4th edition) ✓ Harrison's Principles of Internal Medicine Vol. (19th edition) ✓ Mandell, Douglas & Bennett's Principles and Practice of Infectious Diseases (9th edition) ✓ Ananthanarayan & Panicker's Textbook of Microbiology (10th edition) ✓ Principles & Practice of Clinical Virology (6th edition) 							

Course le	earning outcomes		
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)
Unit 1	Basic anatomy and physiology	Recall basic concepts of human anatomy and physiology. (C1)	4/3 = 7
Unit 2	Principles of Diagnostic Virology	 Illustrate clinical features of viral infections. (C2) Interpret case sheets from hospitals. (C2) Outline differential diagnoses of various clinical presentations. (C2) 	4/3 = 7
Unit 3	Collection, transport and processing of clinical samples	 Demonstrate various sample collection techniques. (C2) Demonstrate sample packaging and transportation techniques. (C2) Demonstrate sample processing techniques. (C2) 	1/4 = 5
Unit 4	Diagnostic algorithms and selection of assays, Disease kinetics, Syndromic approach.	 Summarize diagnostic algorithms of viral infections. (C2) Explain disease kinetics of viral infections. (C2) Explain syndromic approach for diagnosis of viral disease. (C2) 	4/3 = 7
Unit 5	Viral encephalitis / meningitis / meningoencephalitis - Etiology, epidemiology,	 Explain viral encephalitis, meningitis and meningoencephalitis, illustrate their etiologies and clinical features. (C2) 	4/3 = 7



	T .	<u> </u>	T
	laboratory diagnosis, management, prevention and control	 Explain epidemiology and pathogenesis of viruses causing encephalitis. (C2) Summarize diagnostic algorithm for lab diagnosis of viral encephalitis. (C2) Explain the management, prevention and control measures. (C2) 	
Unit 6	Viral diarrhea / Viral food borne illness - Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain viral diarrhea and viral food borne illness, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing diarrhea and food borne illness. (C2) Summarize diagnostic algorithm for lab diagnosis of viral diarrhea and food borne illness. (C2) Explain the management, prevention and control measures. (C2) 	3/3 = 6
Unit 7	Exanthems; Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain exanthems, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing exanthems. (C2) Summarize diagnostic algorithm for lab diagnosis of exanthems. (C2) Explain the management, prevention and control measures. (C2) 	4/3 = 7
Unit 8	Congenital viral infections - Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain congenital viral infections, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing congenital viral infections. (C2) 	3/3 = 6



		 Summarize diagnostic algorithm for lab diagnosis of congenital viral infections. (C2) Explain the management, prevention and control measures. (C2) 	
Unit 9	Viral haemorrhagic fevers - Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain viral haemorrhagic fevers, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing viral haemorrhagic fevers. (C2) Summarize diagnostic algorithm for lab diagnosis of viral haemorrhagic fevers. (C2) Explain the management, prevention and control measures. (C2) 	2/3 = 5
Unit 10	Quality control in Diagnostic Virology laboratory	 Explain quality control measures in diagnostic virology (molecular and serology). (C2) 	1/2 = 3





ass/CGPA 6.5 at UG level (BSc Life es/Health Sciences) le course will familiarise students with lyanced clinical and diagnostic virology.
ass/CGPA 6.5 at UG level (BSc Life es/Health Sciences) ne course will familiarise students with
ass/CGPA 6.5 at UG level (BSc Life es/Health Sciences) ne course will familiarise students with
es/Health Sciences) ne course will familiarise students with
es/Health Sciences) ne course will familiarise students with
es/Health Sciences) ne course will familiarise students with
ferential diagnosis of various clinical esentations. The course will enable students to inderstand the syndromic approach towards agnosis of viral infections and make agnostic algorithms of various viral fections. The course will enable students to perform prious tests required to diagnose various ral infections and selected bacterial fections. The course will make students aware about the reging infectious diseases. The course will learn about the zoonotic fections and concept of one health.
Understand the concepts of clinical and ostic virology. (C1) Interpret & infer information from patient
heets from hospitals. (C2) Outline the clinical features of various
ii hi ha iii hi ha ca la ii



CO 5: Construct diagnostic algorithms of various viral infections. (C6)

CO 6: Demonstrate laboratory skills to perform various diagnostic tests. (C2, P5)

CO 7: Evaluate and discuss case studies in clinical virology. (C5)

Мар	Mapping of COs to POs													
	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	PO1	PO1
COs	1	2	3	4	5	6	7	8	9	10	11	12	3	4
co	✓										\checkmark			
1														
co	✓		✓					✓	✓		✓			
2														
co	✓										\checkmark			
3														
co	✓		✓					✓	✓		\checkmark			
4														
СО	✓		✓	✓				✓	✓		\checkmark			
5														
co	✓		✓	✓		✓			✓		\checkmark		✓	
6														
co	✓		✓	✓				✓	V		✓			
7														

	LEARNING	CONTACT	SLT
	STRATEGY	HOURS	
	Lecture	30	90
	Seminar	12	36
	Small Group	6	18
	Discussion (SGD)		
	Self-directed	4	12
	learning (SDL)		
Learning Strategies, Contact Hours	Problem Based	4	12
ind Student Learning Time (SLT)	Learning (PBL)		
	Case Based Learning	4	12
	(CBL)		
	Clinic	-	-
	Practical	-	-
	Revision	-	-
	Assessment	3	-
	TOTAL	63	180
•	FORMATIVE	SUMMA	ATIVE
Assessment Methods	Assignment	Mid sem	nester exam
Assessment iviethous	Student presentation	End sem	nester exam
	Group discussion		



Mapping of assessment with COs							
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5	CO 6	CO 7
Assignments							✓
Student presentations		✓	✓	✓	✓		✓
Group discussion	✓	✓	✓	✓			✓
Mid-semester examination	✓	✓	✓	✓			
End-semester examination	✓	✓	✓	✓	✓	✓	✓
Practical examination	*	×	×	×	×	×	×

Feedback Methods	Student feedback on Course and Course master.		
Reference Materials	 ✓ Lennette's Laboratory diagnosis of viral infections (4th edition) ✓ Harrison's Principles of Internal Medicine Vol. (19th edition) ✓ Mandell, Douglas & Bennett's Principles and Practice of Infectious Diseases (9th edition) ✓ Ananthanarayan & Panicker's Textbook of Microbiology (10th edition) ✓ Principles & Practice of Clinical Virology (6th edition) 		

Course learning outcomes					
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)		
Unit 1	Viral infections of Respiratory tract- Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain viral infections of respiratory tract, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing infections of respiratory tract. (C2) Summarize diagnostic algorithm for lab diagnosis of viral infections of respiratory tract. (C2) Explain the management, prevention and control measures. (C2) 	4/3 = 7		



		Explain viral STIs, illustrate their	
Unit 2	Viral STIs including HIV: Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain viral STIS, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing STIS. (C2) Summarize diagnostic algorithm for lab diagnosis of viral STIS. (C2) Explain the management, prevention and control measures. (C2) 	4/3 = 7
Unit 3	Human Retroviruses Except HIV- Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain diseases caused by human retroviruses, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of human retroviruses. (C2) Summarize diagnostic algorithm for lab diagnosis of viral infections caused by human retroviruses. (C2) Explain the management, prevention and control measures. (C2) 	5/3 = 8
Unit 4	Viruses and cancer – Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain cancers caused by viruses, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing cancers. (C2) Summarize diagnostic algorithm for lab diagnosis of cancers caused by viruses. (C2) Explain the management, prevention and control measures. (C2) 	1/3 = 4
Unit 5	Slow Viral diseases – Etiology, epidemiology, laboratory diagnosis, management,	 Explain slow viral diseases, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing slow viral diseases. (C2) 	1/3 = 4



	1	1	I
	prevention and control	 Summarize diagnostic algorithm for lab diagnosis of slow viral diseases. (C2) Explain the management, prevention and control measures. (C2) 	
Unit 6	Prion Diseases- Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain prion diseases, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing prion diseases. (C2) Summarize diagnostic algorithm for lab diagnosis of prion diseases. (C2) Explain the management, prevention and control measures. (C2) 	-/3 = 3
Unit 7	Zoonotic Viral infections – Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain zoonotic viral infections, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing zoonotic viral infections. (C2) Summarize diagnostic algorithm for lab diagnosis of viral infections of zoonotic origin. (C2) Explain the management, prevention and control measures. (C2) 	4/3 = 7
Unit 8	Viral Hepatitis	 Explain clinically viral hepatitis, illustrate the etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing hepatitis. (C2) Summarize diagnostic algorithm for lab diagnosis of viral hepatitis. (C2) Explain the management, prevention and control measures. (C2) 	3/3 = 6



Unit 9	Viral Infections in Organ Transplant Recipients and immunocompromi sed individuals	 Explain the viral infections in organ transplant recipients and the immunocompromised, illustrate the etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing transplant related infections and infections in the immunocompromised. (C2) Summarize diagnostic algorithm for lab diagnosis of viruses causing viral infections in organ transplant recipients and the immunocompromised. (C2) Explain the management, prevention and control measures. (C2) 	4/3 = 7
Unit 10	Miscellaneous Viral Infections	 Explain viral infections of eyes, cardiovascular and renal systems, illustrate the etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing infections in eyes, cardiovascular and the renal systems. (C2) Summarize diagnostic algorithm for lab diagnosis of viruses causing infections of the eyes, cardiovascular and renal systems. (C2) Explain the management, prevention and control measures. (C2) 	4/3 = 7





Name of the Program	MSc Clinical Virology
Course Title	Viral Vaccines & Anti-Viral Pharmacotherapy
Course Code	MIV605
Academic Year	2022-2024
Semester	III
Course credits	4
Course Prerequisite	First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences)
Course Synopsis	 The course will offer an overview of viral vaccines. The course will describe the history of vaccine, viral vaccines, types, and newer technologies. The course will offer an overview of antiviral therapeutics. The course will describe the history of antivirals, types and newer technologies in antiviral drug discovery.
Course Outcomes	CO 1: Outline the historical events in vaccine development, basic principles of vaccination and various types of viral vaccines available. (C2) CO 2: Explain the mechanism of action of various viral vaccines and comment on their dosage, side effects, immune response provoked, etc. (C2) CO 3: Explain the newer technologies available, challenges faced in vaccine development, industrial production of vaccines and ways to perform quality check on vaccines. (C2) CO 4: Analyse and interpret scientific journal and learn to present scientific work with an emphasis on public communication skills. (C4) CO 5: Explain the principles of anti-viral therapy and classify the various anti-viral drugs. CO 6: Illustrate the indications, dosage forms, side effects, contraindications etc. of various anti-viral drugs. (C2)



available, challenges faced in anti-viral drug development, manufacturing of drugs and ways to perform quality check. (C2) CO 8: Illustrate anti-viral susceptibility testing methods and explain anti-viral drug resistance. (C2)**Mapping of COs to POs** РО РО PO COs 5 8 9 10 12 13 14 1 2 3 6 11 CO СО **√ √** ✓ **√ √ √ √ √ √ √** CO 3 ✓ ✓ CO 4 СО **√** CO CO 7 **√ √** ✓ **√** CO 8 **LEARNING STRATEGY CONTACT SLT HOURS** 20 60 Lecture Seminar 30 90 **Small Group** 3 9 Discussion (SGD) Self-directed learning 7 21 (SDL) **Learning Strategies, Contact Hours** Problem Based and Student Learning Time (SLT) Learning (PBL) Case Based Learning (CBL) Clinic Practical Revision Assessment 3 **TOTAL** 63 180 **FORMATIVE SUMMATIVE** Assignment Mid semester exam **Assessment Methods** Student presentation End semester exam Group discussion

CO 7: Summarize the newer technologies



Mapping of assessment with COs										
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5	CO 6	CO 7	CO 8		
Assignments			✓		✓		✓	✓		
Student presentations		✓	✓		✓					
Group discussion		✓					✓	✓		
Mid-semester examination	✓	✓			✓	✓	✓			
End-semester examination	✓	✓	✓		✓	✓	✓	✓		
Practical examination	×	×	×	×	×	×	×	×		

Feedback Methods	Student feedback on Course and Course master.					
	Vaccines, 6th & 7th edition - Stanley I	Plotkin, Walter				
	Orenstein, Paul Offit					
Reference Materials	https://www.who.int/topics/vaccines	<u>s/en/</u>				
	https://www.cdc.gov/vaccines/pubs/	pinkbook/prinvac.html				
	https://www.pnas.org/content/pnas/	/111/34/12283.full.pdf				

Course learni	Course learning outcomes							
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)					
Unit 1	History, principles and types of vaccine	 Outline the historical events in vaccine development. (C2) Explain the basic principle of vaccination. (C2) Classify the types of viral vaccines available. (C2) Explain different types of immune response triggered in a host by various types of vaccines. (C2) Summarise the national immunisation schedule India 2020. (C2) 	3/2 = 5					
Unit 2: Viral vaccine-case studies	Measles, Mumps and Rubella	 Outline the history of Measles, Mumps and Rubella vaccines and controversy related to MMR vaccine. (C2) Explain the types of vaccine available for Measles, Mumps and Rubella. (C2) 	2/1 = 3					



Dengue and Zika	 Demonstrate the mechanism of action, the immune response induced, side effects if any and dosage of MMR vaccine respectively. (C2) List the approved vaccines and vaccines under trials for Measles, Mumps and Rubella. (C1) Outline the history of Dengue vaccine and make a note on Philippines event. (C2) Outline the history of Zika virus 	
	 vaccine. (C1) Explain the types of vaccines available for Dengue and Zika. (C2) Demonstrate the mechanism of action, the immune response induced, side effects if any and dosage of Zika and Dengue vaccine respectively. (C2) List the approved vaccines and vaccines under trials for Zika and Dengue. (C1) Explain the challenges faced in development of Dengue vaccine. (C2) 	2/1 = 3
HAV, HBV and HEV	 Outline the history of Hepatitis A Virus (HAV), Hepatitis B Virus (HBV), and Hepatitis E Virus (HEV) vaccine. (C2) Explain the types of vaccines available for HAV, HBV, and HEV. (C2) Demonstrate the mechanism of action, the immune response induced, side effects if any and dosage of HAV, HEV, and HBV vaccine respectively. (C2) 	2/1 = 3



T		
	 List the approved vaccine and vaccines under trials for HAV, 	
	HEV, and HBV. (C1)	
	TIEV, and TIBV. (CI)	
Influenza	Outline the long history of	
and HPV	Influenza vaccines and explain	
	the need for a flu shot every	
	year. (C2)	
	Outline the history of Human	
	Papillomavirus (HPV) vaccine and	
	controversies related to it. (C2)	
	Explain the types of vaccines	1 1/1.
	available for Influenza and HPV.	2/1 = 3
	(C2)Demonstrate the mechanism of	
	action, the immune response	
	induced, side effects if any and	
	dosage for Influenza and HPV	
	vaccine respectively. (C2)	
	 List the approved vaccine and 	
	vaccines under trials for	
	Influenza and HPV. (C1)	
Rota and	Outline the history of Rotavirus	
Rabies	vaccines and controversies	
, (C	related to it. (C2)	
181	Outline the history of Rabies	
	vaccine. (C2)	
	 Explain the types of vaccines 	
5 7	available for Rota and Rabies.	
	(C2)	
	Demonstrate the mechanism of	2/1 = 3
	action, the immune response	2/1 - 3
	induced, side effects if any and dosage for Rota and Rabies	
	vaccine respectively. (C2)	
	 List the approved vaccines and 	
	vaccines under trial for Rota and	
	Rabies. (C1)	
	Summarize the post-exposure	
	prophylaxis of Rabies. (C2)	



KFD and JEV	 Outline the history of Kyasanur Forest Disease (KFD) and Japanese Encephalitis Virus (JEV) vaccines. (C2) Explain the types of vaccines available for KFD and JEV. (C2) Demonstrate the mechanism of action, the immune response induced, side effects if any and dosage of KFD and JEV vaccine respectively. (C2) List the approved vaccines and vaccines under trial for KFD and JEV. (C1) 	2/1 = 3
Polio and VZV	 Outline the history of Polio vaccine and controversies related to it. (C2) Outline the history of Varicella-Zoster Virus vaccine (VZV). (C2) Explain the types of vaccines available for Polio and VZV. (C2) Demonstrate the mechanism of action, the immune response induced, side effects if any and dosage of Polio and VZV vaccines respectively. (C2) List the approved vaccines and vaccines under trial for Polio and VZV. (C1) 	2/1 = 3
Ebola and Smallpox	 Outline the history of Smallpox vaccine. (C1) Outline the history of Ebola virus vaccine. (C1) Explain the types of vaccines available for Smallpox and Ebola virus. (C2) Demonstrate the mechanism of action, the immune response induced, side effects if any and dosage for Ebola and Smallpox vaccines. (C2) 	2/1 = 3



	T		T
		 List the approved vaccines and vaccines under trial for Ebola and Smallpox. (C1) 	
	HIV	 Outline the history of HIV vaccines. (C1) Explain the types of vaccines available for HIV. (C2) Demonstrate the mechanism of action, immune response induced, side effects if any and dosage for HIV vaccine. (C2) List the approved vaccines and vaccines under trial for HIV. (C1) Explain the challenges faced to develop a vaccine for HIV. (C2) 	2/1 = 3
	SARS CoV-2	 Explain the different types of vaccines developed/ being developed for SARS-CoV-2. (C2) Demonstrate the mechanism of action, the immune response induced, side effects if any and dosage for SARS-CoV-2 vaccine. (C2) List the vaccines that have completed the clinical trials. (C1) 	2/1 = 3
Unit 3	Newer technologies, challenges and vaccine trial	 Demonstrate the different new technologies that have been used in recent years or the ones that can be used for vaccine development. (C2) Explain the challenges faced in the development of a vaccine. (C2) Outline the various steps involved in vaccine trials. (C2) Explain in brief about each phase involved in vaccine trials. (C2) 	2/1 = 3
Unit 4	Industrial production and quality	Demonstrate the process of industrial production of vaccines. (C2)	2/1 = 3



	ala al c	- 1	<u> </u>
	checking of vaccines	 Explain in brief the different techniques used at various stages of vaccine production for conducting quality check. (C2) 	
Unit 5	Introduction and Antiviral drugs - classification	 Outline concept of antiviral therapy and historical perspectives. (C2) Translate process of antiviral drug development. (C2) Classify antiviral drugs based upon their site of attack on viral replication cycle. (C2) 	3/2 = 5
Unit 6	Mechanism of action, therapeutic indications, adverse side effects of anti-viral drugs and potential drug interactions	 Explain mechanism and therapeutic actions of different antiviral drugs. (C2) Illustrate different antiviral dosage forms. (C2) Summarize adverse side effects, potential drug interactions and contraindications. (C2) 	3/2 = 5
Unit 7	Antiviral susceptibility testing methods and Antiviral drug resistance	 Describe the different antiviral susceptibility testing methods. (C2) Explain antiviral drug resistance. (C2) Illustrate the factors contributing to antiviral drug resistance. (C2) 	3/2 = 6
Unit 8	Emerging trends, challenges and prospects in antiviral research with a focus on ongoing antiviral drug trials and newer drugs in pipeline	 Summarize trends, challenges and prospects in antiviral drug discovery and development. (C2) Outline antiviral drug trials and drugs in pipeline. (C2) 	2/1 = 3





Manipal Institute of Virology

CURRICULUM

Name of the Program	MSc Clinical Virology
Course Title	Insect vectors of Viral diseases
Course Code	MIV607
Academic Year	2022-2024
Semester	III
Course credits	2
Course Prerequisite	First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences)
	 To understand the basic concept of medical entomology, vector-borne disease (VBD) epidemiology and biology of arthropods. To understand the important VBD's
Course Synopsis	transmission cycles, arthropod adaptation and diagnostic tools for VBD.
Course Synopsis	To understand the taxonomy, surveillance methods and control programmes of
	important vectors.4. To understand the impact of climate and environment on proliferation of vectors and spread of VBDs.
Albill like	CO 1: Outline the basic terminologies of medical entomology and epidemiology. (C2) CO 2: Explain biology and ecology of important arthropods. (C2) CO 3: Explain the transmission cycle of important VBDs of India and its host adaptations. (C2)
Course Outcomes	CO 4: Outline the molecular and immunological tools for VBD detection in vectors. (C2) CO 5: Explain the taxonomic classification and morphological characteristics of arthropods. (C2) CO 6: Explain surveillance tools and control strategies of important public health vectors. (C2)



								CO 7: Outline the impact of climate and						
							envi	ronm	ent oi	า vecto	r ecolo	gy. (C2	2)	
Мар	ping c	of COs	to PO	Os										
	РО	РО	РО	РО	РО	РО	РО	РО	РО	PO1	РО	РО	PO	PO1
COs	1	2	3	4	5	6	7	8	9	0	11	12	13	4
CO 1	✓										✓			
CO 2	✓										✓			
CO 3	✓		✓			✓		✓			✓			
CO 4	✓					✓					✓			
CO 5	✓										✓			
CO 6	✓		✓			✓					✓			
CO 7	✓		√					✓			√			

	LEARNING STRATEGY	CONTACT HOURS	SLT	
	Lecture	10	30	
	Seminar	4	12	
	Small Group Discussion (SGD)	1	3	
Learning Strategies, Contact Hours and Student Learning Time (SLT)	Self-directed learning (SDL)	-	-	
	Problem Based			
	Case Based Learning (CBL)	-	-	
	Clinic	-	-	
	Practical	30	90	
	Revision	-	-	
	Assessment	1	-	
	TOTAL	46	135	
	FORMATIVE	SUMMATIN	/E	
Anna and Marka da	Assignment	Mid semester exam		
Assessment Methods	Student presentation			
	Group discussion			

Mapping of assessment with COs								
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5	CO 6	CO 7	
Assignments		✓	✓			✓		
Student presentations		✓	✓	✓	✓	✓	✓	
Group discussion							✓	
Mid-semester examination	✓	✓	✓					
End-semester examination	×	×	×	×	×	×	×	
Practical examination (Internal Assessment)				✓	✓	✓		



Feedback Methods	Student feedback on Course and Course master.				
Reference Materials	 Medical entomology for students – Mike William Service Vector control: Methods for Use by Individuals and Communities – Jan A. Rozendaal Guide to Entomology – Mike William Service Medical Entomology: A Textbook on Public Health and Veterinary Problems Caused by Arthropods- B F Eldridge 				

Course le	Course learning outcomes							
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials + Practicals)					
Unit 1	Introduction to Medical Entomology	 Outline the basic definitions and terminologies of medical Entomology and Epidemiology. (C2) Explain the role of vectors in epidemiological triad. (C2) List globally known arthropod vectors. (C1) Classify different modes of vector-borne disease (VBD) transmission. (C2) 	2					
Unit 2	Biology and ecology of medical important Arthropods	 Explain the Life cycle, host seeking behaviour, resting behaviour, feeding behaviour, breeding Habitat types and Oviposition behaviour of Diptera (Mosquito, sandfly, tsetse fly, black fly and triatomine bug). (C2) Explain the Life cycle, host seeking behaviour, resting behaviour, feeding behaviour, breeding Habitat types and Oviposition behaviour of Arachnids (Ticks and mites). (C2) Explain the Life cycle, host seeking behaviour, resting behaviour, feeding behaviour, breeding Habitat types and Oviposition 	2+4 = 6					



	1		
		behaviour of Siphonaptera. (Fleas)	
		(C2)	
Unit 3	Common vector borne diseases	 Outline the global and national Importance of VBDs. (C2) Explain the transmission cycle, symptoms and treatment of Parasite origin VBDs (Malaria, Filariasis and Leishmaniasis). (C2) Explain the transmission cycle, symptoms, treatment, evolution and mutation of Arboviral origin VBDs (Dengue, Chikungunya, Japanese encephalitis, Kyasanur Forest Disease (KFD), West Nile, Crimean Congo Haemorrhagic fever (CCHF) and Yellow fever). (C2) Explain the transmission cycle, symptoms and treatment of Bacterial and rickettsial diseases. (C2) Outline the Interaction of Arboviruses with various hosts. (C2) 	2+2 = 4
Unit 4	Vector control programmes	 Outline the National Vector Borne-Disease Control Programmes of India. (C2) Explain the Malaria, Filaria and Kala-azar control strategies of India. (C2) Explain the Arboviral control strategies of India. (C2) 	2+1 = 3



	Τ		
Unit 5	Vector surveillance	 Outline the arthropod Sampling methods. (C2) List the commonly used entomological measures. (C1) Outline the collection tools for various medically important arthropods (Mosquitoes, ticks, mites, sand flies, flea and other arthropods). (C2) Demonstrate Adult mosquito collection tool. (C2) Survey Mosquito Immature. (C4) Demonstrate tick collection technique. (C2) Explain the port surveillance and Xeno-monitoring. (C2) Explain the VBD outbreak Investigation. (C2) Utilize the Geographical information system (GIS) in vector Surveillance and control. (C3) 	2+2 = 4
Unit 6	Arthropod	 Classify the taxonomy of Arthropoda. (C2) Demonstrate morphological characterization of medically important arthropod species in India. (C2) Identify morphology of Aedes mosquito. (C3) Identify morphology of Culex mosquito. (C3) Identify morphology of Ixodid ticks. (C3) Explain molecular taxonomy and preservation techniques of arthropods. (C2) 	2+10 = 12
Unit 7	Vector control	Outline the principles and types of vector control (Mechanical, Chemical, Biological,	1+1 = 2



	1		T .
Unit 8	Diagnostics of vector-borne diseases	 Environmental management and Transgenic insect technique). (C2) Explain the Integrated Vector Management (IVM), Insecticide resistance and management. (C2) Explain personal protective measures against vectors. (C2) Demonstrate molecular diagnosis of VBDs (PCR and Sequencing). (C2) Demonstrate Insect sample preparation and processing for molecular detection. (C2) Demonstrate VBD immunodiagnosis (ELISA, ICT, IFA and 	2+5 = 7
		 Neutralisation). (C2) Explain the inter-seasonal maintenance of arboviral diseases. (C2) Explain the impact of climatic changes on vector biodiversity and migration. (C2) 	
Unit 9	Impact of Environmental and Climatic on Vector ecology	 Explain the impact of Pollution and Urbanization on Vector proliferation. (C2) Outline Emerging and re-emerging vector-borne diseases. (C2) Outline the notifiable diseases and Public health emergencies of international concern (PHEIC). (C2) 	1+4 = 5





Name	of th	e Pro	gram				MSc	Clinic	cal Vir	ology						
Cours	e Titl	е			Virology lab design and management											
Course Code							MIV609									
Academic Year							202	2-202	4							
Semester III																
Cours	e cre	dits					1					\ ^				
Cours	o Dro	rogui	cito				First	class	/CGP/	4 6.5 a	t UG	lev	el (BSc	Lif	е	
Cours	ePre	requi	site				Scie	nces/	Health	n Scien	ces)					
							This	cour	se inte	ends to	aco	quai	nt stu	den	ts v	vith the
Cours	o Svn	oncic					min	imum	requi	remen	ts fo	or es	stablis	hing	gar	national
Cours	e Syn	opsis					viro	logy l	abora	tory, k	кеер	ing	in vie	w t	he	
							eme	ergen	ce of n	iew vir	al pa	athc	gens.			
							CO :	1: Illus	strate	key el	eme	nts	of a vii	rolo	gy	
									y. (C2)							
Cours	e Out	tcome	es							equirer			desig	ning	gа	
							_			ogy lak	•	•				
							l l			uality s	syste	ems	applic	able	e in	а
Man	nina	of CO	c to D	06	\rightarrow		uiag	nosti	c lab.	(C2)						
Iviap	PO	PO	s to P	PO	PO	РО	РО	РО	PO	РО	PC)	РО	PC		РО
COs	1	2	3	4	5	6	7	8	9	10	11		12	13		14
CO 1	✓		~	✓	✓		✓	✓			,	/				
CO 2	1		1			✓		✓	✓		,	/				
CO 3	~		✓		✓				✓		✓					
	113	1				1	LE	ARNII	NG ST	RATEG	Υ	СС	NTAC	Т		SLT
												Н	OURS			
							Lec	cture					7			21
Learning Strategies, Contact Hours					Seminar					5			15			
and S	and Student Learning Time (SLT)					Sm	all Gr	oup				1			3	
					Dis	cussi	on (SG	iD)								
						Sel	f-dire	cted le	earnin	g		1			3	
							(SE	DL)								



	Problem Based	1	3
	Learning (PBL)		
	Case Based Learning	-	-
	(CBL)		
	Clinic	-	-
	Practical	ı	-
	Revision	-	-
	Assessment	1	
	TOTAL	16	45
	FORMATIVE	SUMMAT	IVE
Assessment Methods	Assignment	Mid seme	ster exam
Assessment Methods	Student presentation		
	Group discussion		

Mapping of assessment with course outcomes						
Nature of assessment	CO 1	CO 2	CO 3			
Assignments		✓				
Student presentations	√					
Group discussion	✓	✓	✓			
Mid-semester examination	✓	✓	✓			
End-semester examination	×	×	×			
Practical examination	×	×	×			

Feedback Methods	Student feedback on Course and Course master.				
Reference Materials	 Guidelines on establishment of virology laboratory in developing countries Guideline Document for design of BSL-2 Labs (District Hospitals, CHC And PHC Level) 				

Course learning outcomes							
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)				
Unit 1	Introduction to virology lab	 Explain key components of a virology lab. (C2) Compare and contrast requirements of a diagnostic and research virology lab. (C2) 	3/2 = 5				



		 Summarize biosafety principles. (C2) Outline primary barriers of biosafety. (C2) Illustrate requirements of facility design and construction. (C2) Outline requirements for different biosafety level laboratories. (C2) 	
Unit 2	Virology lab design	 Develop a model of virology lab with BSL-2 facility. (C3) Develop a model of virology lab with BSL-3 facility. (C3) 	3/2 = 5
Unit 3	Quality systems in a diagnostic virology lab	 Outline components of quality management. (C2) Explain concepts of preanalytical, analytical and post analytical quality parameters. (C2) Summarize lab accreditation guidelines/policies. (C2) 	3/2 = 5





Name of the Program	MSc Clinical Virology			
Course Title	Intellectual Property rights and patenting			
Course Code	MIV611			
Academic Year	2022-2024 III First class/CGPA 6.5 at UG level (BSc Life Sciences/Health Sciences) 1. This course will sensitize students about the basic information on Intellectual Propert Rights, Patent filing process, analysing and drafting patent applications, monetizing research results. 2. The course will help students to appreciate the need for various kinds of intellectual propert (IP) protection and its impact and association with the growth of organisations. 3. The course will teach students to appreciate the students to appreciate with the growth of organisations.			
Semester	III			
Course credits	1			
Course Prerequisite				
Course Synopsis Course Outcomes	 This course will sensitize students about the basic information on Intellectual Property Rights, Patent filing process, analysing and drafting patent applications, monetizing research results. The course will help students to appreciate the need for various kinds of intellectual property (IP) protection and its impact and association with the growth of organisations. 			
	principles and procedure for obtaining patent. (C4) CO 3: Apply technical concepts of IP related			
	technology. (C3)			



CO 4: Demonstrate and develop awareness of relevance and impact of intellectual property law on academic and professional lives. (C3) **CO 5:** Analyse ethical and professional issues which arise in intellectual property law context. (C4)**Mapping of COs to POs** PO COs 1 2 3 5 8 10 11 12 13 14 7 CO 1 CO ✓ 2 CO ✓ СО co

Learning Strategies, Contact Hours
and Student Learning Time (SLT)

LEARNING STRATEGY	CONTACT	SLT
	HOURS	
Lecture	7	21
Seminar	5	15
Small Group Discussion	1	3
(SGD)		
Self-directed learning	2	6
(SDL)		
Problem Based Learning	-	-
(PBL)		
Case Based Learning	-	-
(CBL)		
Clinic	-	-
Practical	-	-
Revision	-	-
Assessment	1	-
TOTAL	16	45
FORMATIVE	SHMMATI	/F

Assessment Methods

5

FORMATIVE	SUMMATIVE	
Assignment	Mid semester exam	
Student presentation		
Group discussion		



Mapping of assessment with COs					
Nature of assessment	CO 1	CO 2	CO 3	CO 4	CO 5
Assignments	✓				
Student presentations				✓	
Group discussion					
Mid-semester examination	✓	✓	✓	✓	✓
End-semester examination	*	*	×	×	×
Practical examination	×	*	*	×	×

Feedback Methods	Student feedback on Course and Course master.
Reference Materials	 Intellectual Property Rights, Deborah. E. Bouchoux, Cengage Learing. Intellectual Property Rights – Unleashmy The Knowledge Economy, Prabuddha Ganguli, Tate Mc Graw Hill Publishing Company Ltd.

Course le	arning outcom	es	
Content	Topics	Learning Outcomes	Hours (Lectures/Tutorials)
Unit 1	Overview of Intellectual Property	 Explain intellectual property and its importance. (C2) Classify the various types of intellectual property. (C2) List the various international organizations associated with IPR. (C2) List the various agencies and treaties associated with IPR. (C2) Explain the importance of intellectual property rights. (C2) 	1/2 = 3
Unit 2	Patents	 Explain the definition, basics of patents. (C2) Outline the patent searching process. (C2) Reading and interpreting patent documents. (C2) Summarize in detail the ownership rights and transfer. (C2) 	2/2 = 4



Explain the new developments in	
Patent law. (C2)	
Outline the International Patent	
Law. (C2)	
Discuss important case laws. (C3)	
Explain Fundamental of Copy Right	
Law. (C2)	
Explain the importance of	
originality of material in Law of	
copy rights. (C2)	
Explain the rules of rights of	
Unit 3 Copyrights reproduction. (C2)	2/2 - 4
Explain the rights to perform the	2/2 = 4
work publicly. (C2)	
Explain the issues of copyrighted	
owner. (C2)	
Explain the copyright law globally.	
(C2)	
Discuss important case laws. (C3)	
Define trademarks and explain their	
purpose and function. (C2)	
Illustrate importance of acquisition	
of trademark rights. (C2)	
Explain the trademark protectable	
Unit 4 Trademarks matter topics. (C2)	2/2 = 4
Explain the selection and evaluation	
of a trademark. (C2)	
Explain the trademark registration	
process. (C2)	
Discuss important case laws. (C3)	





Name of the Program	MSc Clinical Virology
Course Title	Comprehensive Practical (Clinical & Diagnostic
Course Title	Virology and Laboratory rotation -II)
Course Code	MIV613
Academic Year	2022-2024
Semester	III
Course credits	4
Course Provoquisite	First class/CGPA 6.5 at UG level (BSc Life
Course Prerequisite	Sciences/Health Sciences)
Course Synopsis	Refer to MIV601, MIV603 and MIV512
	CO 1: Analyse clinical cases and design diagnostic
	algorithms. (C4)
	CO 2: Demonstrate laboratory skills to perform
	various diagnostic tests. (P5)
	CO 4: Understand the syndromic approach of
Cavina Outagnes	finding the etiological agents. (C2)
Course Outcomes	CO 5: Understand the choice of tests depending on
	different patient parameters. (C2)
	CO 6: Correlate the clinical picture with the
	laboratory findings. (C4)
	CO 7: Understand appropriate reporting of test
	results. (C3)

Map	ping	of CO	s to P	Os										
	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО
COs	1	2	3	4	5	6	7	8	9	10	11	12	13	14
CO 1	~		√	√		√			✓		√		✓	
CO 2	✓		√	✓		√			✓		✓		✓	
CO	✓										✓			
CO 4	✓		√					√	✓		✓			
CO 5	√		√	√				√	√		√			
CO 6	✓		√	✓		√			√		√		√	



CO 7	✓		✓	✓				✓	✓		√					
							LEA	RNING	STR.	ATEGY		NTACT OURS	S	SLT		
							Lecti	ıre				-		-		
							Semi	inar				-		-		
							Smal	l Grou	р			-		-		
							Discu	ussion	(SGD)						
							Self-	direct	ed lea	rning		-		-		
							(SDL))								
Learn	_	_					Prob	lem B	ased			-		-		
and S	tuder	nt Lea	rning	Time	(SLT)		Learı	ning (F	PBL)							
							Case	Based	d Lear	ning		- /		-		
							(CBL))								
							Clinic	2				7.		-		
							Pract	tical				120	3	860		
							Revis	sion						-		
							Asse	ssmer	nt			2		-		
							TOTA	4L				122	3	860		
Accor	smen	+ Mai	hode				FORI	MATI\	/E		9	SUMMA [*]	TIVE			
Asses	sillen	ı ivie	uious				Inter	nal as	sessn	nent	E	nd seme	ester e	xam		

Feedback Methods	Student feedback on Course and Course master.
Reference Materials	Refer to MIV601, MIV603 and MIV512

Course lea	Course learning outcomes											
Content	Topics	Learning Outcomes	Hours (Practicals)									
Unit 2 (MIV601)	Principles of Diagnostic Virology	 Illustrate clinical features of viral infections. (C2) Interpret case sheets from hospitals. (C2) Outline differential diagnoses of various clinical presentations. (C2) 	3									
Unit 3 (MIV601)	Collection, transport and processing of clinical samples	Demonstrate various sample collection techniques. (C2)	3									



		 Demonstrate sample packaging and transportation techniques. (C2) Demonstrate sample processing techniques. (C2) 	
Unit 5 (MIV601)	Viral encephalitis / meningitis / meningoencephalitis - Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain viral encephalitis, meningitis and meningoencephalitis, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing encephalitis. (C2) Summarize diagnostic algorithm for lab diagnosis of viral encephalitis. (C2) Explain the management, prevention and control measures. (C2) 	4
Unit 6 (MIV601)	Viral diarrhea / Viral food borne illness - Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain viral diarrhea and Viral food borne illness, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing diarrhea and food borne illness. (C2) Summarize diagnostic algorithm for lab diagnosis of viral diarrhea and food borne illness. (C2) Explain the management, prevention and control measures. (C2) 	4
Unit 7 (MIV601)	Exanthems; Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain exanthems, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing exanthems. (C2) Summarize diagnostic algorithm for lab diagnosis of exanthems. (C2) 	4



	T		T
		 Explain the management, prevention and control measures. (C2) 	
Unit 8 (MIV601)	Ricketssial Diseases- Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain Ricketssial diseases, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing Ricketssial diseases. (C2) Summarize diagnostic algorithm for lab diagnosis of Ricketssial diseases. (C2) Explain the management, prevention and control measures. (C2) 	4
Unit 9 (MIV601)	Congenital viral infections - Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain congenital viral infections, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing congenital viral infections. (C2) Summarize diagnostic algorithm for lab diagnosis of congenital viral infections. (C2) Explain the management, prevention and control measures. (C2) 	4
Unit 10 (MIV601)	Viral haemorrhagic fevers - Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain viral haemorrhagic fevers, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing viral haemorrhagic fevers. (C2) Summarize diagnostic algorithm for lab diagnosis of viral haemorrhagic fevers. (C2) Explain the management, prevention and control measures. (C2) 	4



Content	Topics	Learning Outcomes	Hours (Practicals)
Unit 1 (MIV603)	Viral infections of Respiratory tract- Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain viral infections of respiratory tract, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing infections of respiratory tract. (C2) Summarize diagnostic algorithm for lab diagnosis of viral infections of respiratory tract. (C2) Explain the management, prevention and control measures. (C2) 	5
Unit 2 (MIV603)	Viral STIs including HIV; Chlamydia - Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain viral STIs, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing STIs. (C2) Summarize diagnostic algorithm for lab diagnosis of viral STIs. (C2) Explain the management, prevention and control measures. (C2) 	5
Unit 3 (MIV603)	Human Retroviruses Except HIV- Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain diseases caused by human retroviruses, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of human retroviruses. (C2) Summarize diagnostic algorithm for lab diagnosis of viral infections caused by human retroviruses. (C2) Explain the management, prevention and control measures. (C2) 	4



Unit 4 (MIV603)	Viruses and cancer - Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain cancers caused by viruses, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing cancers. (C2) Summarize diagnostic algorithm for lab diagnosis of cancers caused by viruses. (C2) Explain the management, prevention and control measures. (C2) 	4
Unit 5 (MIV603)	Slow Viral diseases - Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain slow viral diseases, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing slow viral diseases. (C2) Summarize diagnostic algorithm for lab diagnosis of slow viral diseases. (C2) Explain the management, prevention and control measures. (C2) 	4
Unit 6 (MIV603)	Prion Diseases- Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain prion diseases, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing prion diseases. (C2) Summarize diagnostic algorithm for lab diagnosis of prion diseases. (C2) Explain the management, prevention and control measures. (C2) 	4
Unit 7 (MIV603)	Zoonotic Viral infections - Etiology, epidemiology, laboratory diagnosis, management, prevention and control	 Explain zoonotic viral infections, illustrate their etiologies and clinical features. (C2) Explain epidemiology and pathogenesis of viruses causing zoonotic viral infections. (C2) 	4



	Summarize diagnostic algorithm	
	for lab diagnosis of viral	
	infections of zoonotic origin. (C2)	
	 Explain the management, 	
	prevention and control measures.	
	(C2)	

Laborato	ory Rotation -II		
Posting	Sections	Learning Outcomes	Hours
1	Serology	Observe and understand the workflow in the diagnostic laboratory.	60 hours (12 hours in each section)
2	Sample processing and Extraction	Observe, understand and develop skills of processing clinical samples.	,
3	Molecular Diagnostics	Observe and understand the principle, methodology and technique of the different tests.	
4	Tissue culture	technique of the different tests (serological, molecular and cell culture based) performed routinely	
5	Sample reception, Barcoding, Sample Storage and Decontamination	 in a diagnostic laboratory. Develop skills to perform diagnostic tests independently. Perform sample reception independently, and observe barcoding and sample storage processes. Observe, understand and perform the protocols of decontamination and discarding of biomedical waste. 	



FOURTH SEMESTER





Manipal Institute of Virology

CURRICULUM

Name	of th	e Pro	gram					MSc (Clinica	al Virol	ogy						
Course	e Title	9						Research Project									
Course	e Cod	е						MIV699									
Acade	mic Y	'ear						2022-2024									
Semes	ster							IV									
No. of	cred	its						18)				
Course	e Prei	requi	site					First	class/	CGPA	6.5 at	UG I	evel (B	Sc Life			
								Scien	ces/H	ealth S	cience	s)					
Course Synopsis									rry ou weel e prostituti any ist wi	ut a Proks. oject ron/ in other th app	nay be dustry/ institu	ork for e carri reseation we f the pa	a mini ed out rch lab where f	in the coratory facilities estitute.			
Cours	Course Outcome							CO2: (C4) CO3: study CO4: (C4) CO5: the e CO6: (C4, F CO7: exam CO8:	Delin Procu Devis xperir Analy Prese iners. Trans	eate ai re reque e the rements. rse and ent rese (C5, P	ms and uired method (C5, P3 docunes) documents (C5, A5) are scie	litera l object nateria ology nent th	and exe ne outco to the p	f the e work.			
Мар	ping o	of CO	s to P	Os							<u> </u>						
	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО	РО			
COs	1	2	3	4	5	6	7	8	9	10	11	12	13	14			
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5 CO														
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CO 7	✓	✓	✓	✓	✓	✓		✓	✓	✓	✓	✓	✓	
CO 8	√	√	√	√	√	✓		✓	√	√	✓	✓	~	
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8. PROGRAM OUTCOMES & COURSE OUTCOMES MAPPING

S. No.	Course Code	Course Name	Credits	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO1 0	PO11	PO1 2	PO1 3	PO14
1.	MIV501	Cell Biology	1	CO1 CO2 CO3 CO4 CO5 CO6 CO7							CO1 CO2 CO5 CO7	CO6 CO8		CO1 CO2 CO3 CO4 CO5 CO6 CO7 CO8			
2.	MIV503	Basic Virology	3	CO1 CO2 CO3 CO4 CO5 CO6 CO7		CO4 CO5		CO6 CO7	CO6 CO7			CO4		CO1 CO2 CO3 CO4 CO5 CO6 CO7			
3.	MIV505	Biosafety, Biosecurity and Bioethics	3	CO1 CO2 CO3 CO4 CO5 CO6 CO7 CO8 CO9 CO10 CO11	CO3 CO4 CO5 CO1 1	CO1 CO2 CO4 CO5 CO6 CO11 CO12	CO4 CO6	CO1 1	CO2 CO3 CO4 CO6 CO9 CO10 CO11	CO1 1	CO1 CO9 CO10 CO11	CO1 CO10 CO11 CO12	CO1 2	CO1 CO2 CO3 CO4 CO5 CO6 CO7 CO8 CO9 CO10 CO11 CO12	CO7 CO8 CO9 CO1 0 CO1 1 CO1 2		CO7
4.	MIV507	Tissue/Cell culture	2	CO1 CO2 CO3 CO4 CO5		CO3 CO4		CO4	CO2 CO3		CO1	CO2		CO1 CO2 CO3 CO4 CO5		CO3	
5.	MIV509	Systematic Virology	6	CO1 CO2 CO3 CO4	CO1 CO3	CO1 CO3 CO4		CO1			CO3 CO4			CO1 CO2 CO3 CO4			

6.	MIV511	Immunology of Viral diseases	3	CO1 CO2 CO3 CO4 CO5 CO6 CO7 CO8 CO9 CO10		CO2 CO3 CO6 CO9 CO10 CO11					CO3 CO5 CO9 CO10 CO11	CO10		CO1 CO2 CO3 CO4 CO5 CO6 CO7 CO8 CO9 CO10			
7.	MIV513	Practical I	1	CO1 CO2 CO3		CO2			CO1 CO2			CO1		CO1 CO2 CO3		CO2	
8.	MIV515	Microbiology posting	2	CO1 CO2 CO3		CO2			CO1 CO2			CO1		CO1 CO2 CO3		CO2	
9.	MIV502	Epidemiology	3	CO1 CO2 CO3 CO4 CO5 CO6	CO3 CO4 CO5	CO1 CO2 CO4 CO5 CO6 CO7	CO4 CO5 CO6	CO6		CO4 CO5	CO2	CO4 CO7	CO5 CO6	CO1 CO2 CO3 CO4 CO5 CO6 CO7	CO4 CO5		CO4 CO5
10.	MIV504	Molecular Virology and Bioinformatics	4	CO1 CO2 CO3 CO4 CO5 CO6	CO2	CO2 CO3 CO5 CO6	CO2 CO4 CO5 CO6	CO2 CO3 CO5 CO6	CO3 CO4 CO5 CO6		CO5 CO6 CO7	CO2	CO2 CO3 CO4 CO5 CO6	CO1 CO2 CO3 CO4 CO5 CO6			
11.	MIV506	Virological Techniques	4	CO1 CO2 CO3 CO4 CO5 CO6 CO7 CO8		CO3 CO7 CO8 CO9	CO6 CO7 CO8 CO9	CO6 CO7 CO8 CO9	CO1 CO2 CO3 CO4 CO5 CO6 CO7 CO8 CO9		CO1 CO7 CO8 CO9	CO5	CO 7 CO8 CO9	CO1 CO2 CO3 CO4 CO5 CO6 CO7 CO8 CO9		CO1 CO3	
12.	MIV508	Analytical Tools (Application of GIS and Biostatistics)	2	CO1 CO2 CO3 CO4 CO5 CO6		CO2 CO3 CO5 CO9 CO11 CO12	CO2 CO4 CO7 CO8 CO9	CO8 CO9 CO1 0 CO1	CO3 CO5		CO8	CO3 CO4 CO5 CO8 CO9	CO1 CO2 CO3 CO4 CO5 CO7	CO1 CO2 CO3 CO4 CO5 CO6	CO6 CO7 CO8 CO9 CO1	CO1	

				CO7 CO8 CO9 CO10 CO11 CO12		CO2	CO1 0 CO1 1 CO1 2	CO1 2			CO1	CO12	CO8 CO9 CO1 0 CO1 1 CO1 2	CO7 CO8 CO9 CO10 CO11 CO12	CO1 1 CO1 2	CO4	CO4
13.	MIVEL510.1	Emerging Viral Diseases and Public health response	3	CO2 CO3 CO4 CO5							CO2 CO4	CO4		CO2 CO3 CO4 CO5			
14.	MIVEL510.2	One health approach in Virology	3	CO1 CO2 CO3 CO4 CO5 CO6		CO2 CO4 CO5 CO6	CO5 CO6		CO6		CO2 CO4 CO5	CO2 CO4 CO5 CO6		CO1 CO2 CO3 CO4 CO5 CO6	CO3	CO6	
15.	MIV512	Practical II	3	CO1 CO2 CO3 CO4 CO5 CO6 CO7	CO2	CO2 CO3	CO2 CO7	CO2 CO3 CO7	CO3 CO4 CO5 CO6 CO7		CO4	CO2 CO6	CO2 CO3	CO1 CO2 CO3 CO4 CO5 CO6 CO7		CO4	
16.	MIV514	Laboratory Rotation - I	2	CO1 CO2		CO2			CO1 CO2			CO1		CO1 CO2		CO2	
17.	MIV601	Clinical & Diagnostic Virology-I	4	CO1 CO2 CO3 CO4 CO5 CO6 CO7		CO2 CO4 CO5 CO6 CO7	CO5 CO6 CO7		CO6		CO2 CO4 CO5 CO7	CO2 CO4 CO5 CO6 CO7		CO1 CO2 CO3 CO4 CO5 CO6 CO7		CO6	
18.	MIV603	Clinical & Diagnostic Virology-II	4	CO1 CO2 CO3 CO4 CO5 CO6 CO7		CO2 CO4 CO5 CO6 CO7	CO5 CO6 CO7		CO6		CO2 CO4 CO5 CO7	CO2 CO4 CO5 CO6 CO7		CO1 CO2 CO3 CO4 CO5 CO6		CO6	
19.	MIV605	Viral Vaccines & Antiviral Pharmacotherapy	4	CO1 CO2	CO4 CO5 CO6	CO2 CO3		CO7	CO2 CO3 CO7	CO7 CO8	CO2 CO3 CO4	CO2 CO4 CO5		CO1 CO2		CO3 CO7	

				CO3 CO4 CO5 CO6 CO7	CO7 CO8				CO8		CO7 CO8	CO6 CO7 CO8		CO3 CO4 CO5 CO6 CO7 CO8			
20. MIN	V607	Insect vectors of Viral diseases	2	CO1 CO2 CO3 CO4 CO5 CO6 CO7	٠	CO3 CO6 CO7			CO3 CO4 CO6		CO3 CO7			CO1 CO2 CO3 CO4 CO5 CO6 CO7			
21. MIN	V609	Virology lab design and management	1	CO1 CO2 CO3		CO1 CO2 CO3	CO1	CO1 CO3	CO2	CO1	CO1 CO2	CO2 CO3		CO1 CO2 CO3			
22. MIN	V611	Intellectual Property rights and patenting	1	CO1 CO2 CO3 CO4	CO1 CO4 CO5	CO1 CO2 CO3 CO4				CO1	CO2 CO3 CO5	CO2 CO3 CO4 CO5		CO1 CO2 CO3 CO4 CO5	CO4 CO5	CO2 CO4	CO4
23. MI\	V613	Comprehensive Practical	4	CO1 CO2 CO3 CO4 CO5 CO6		CO1 CO2 CO4 CO5 CO6 CO7	CO1 CO2 CO5 CO6 CO7		CO1 CO2 CO6		CO4 CO5 CO7	CO1 CO2 CO4 CO5 CO6 CO7		CO1 CO2 CO3 CO4 CO5 CO6 CO7		CO1 CO2 CO6	
24. MIN	V699	Research Project	18	CO1 CO2 CO7 CO8	CO4 CO7 CO8	CO1 CO3 CO5 CO6 CO7 CO8	CO5 CO6 CO7 CO8	CO3 CO5 CO6 CO7 CO8	CO3 CO5 CO7 CO8	CO5	CO1 CO3 CO5 CO6 CO7 CO8	CO5 CO7 CO8	CO2 CO6 CO7 CO8	CO2 CO3 CO6 CO7 CO8	CO5 CO6 CO7 CO8	CO2 CO4 CO5 CO7 CO8	

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